Candidate MRTP Aerosol has Low Impact on Organotypic Gingival Cultures^[1]



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SUMMARY

Introduction. Smoking is one of the major lifestyle-related risk factors for periodontal diseases. Smoking can affect the structure of the mucosa, impair the inflammatory epithelial response, and change the redox status of the cavity. Harm reduction through the oral development of Modified Risk Tobacco Products (MRTP) provides a promising opportunity for adult smokers who would otherwise continue cigarette smoking. An MRTP is defined by the U.S. Family Smoking Prevention and Tobacco Control Act as "any tobacco product that is sold or distributed for use to reduce harm or the risk of tohacco related disease associated with

Human gingival epithelial organotypic cultures. EpiGingival[™] (MatTek corp., Ashland USA) derived from a 46 year old male donor, non-smoker.

Histological analysis. Tissue sections were stained with Hematoxylin (HE). For & Eosin immunohistochemical slides staining, the were incubated with an E-cadherin antibody (Leica Biosystem PA0387, undiluted) and counterstained with hematoxylin.

Adenylate Kinase (AK)-based cytotoxicity. The activity of AK was measured in the basolateral medium using the ToxiLight[™] bioassay kit (Lonza, Rockland, MA, USA).



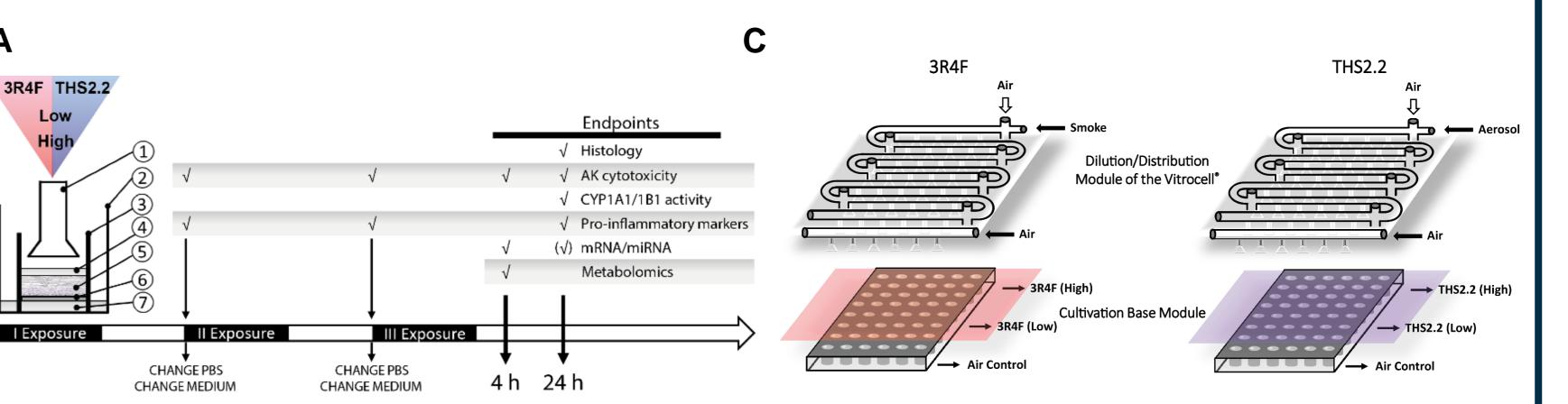
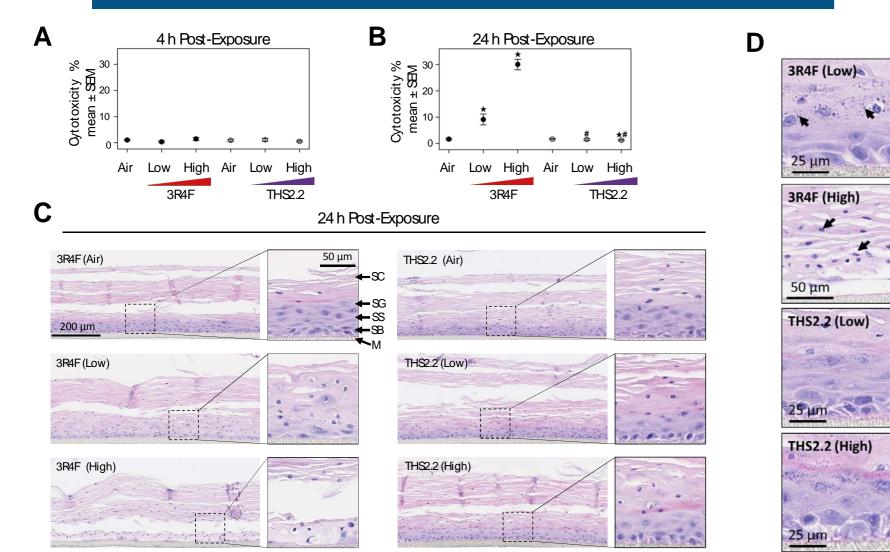


Figure 1. Study design and exposure system. (A) Human gingival

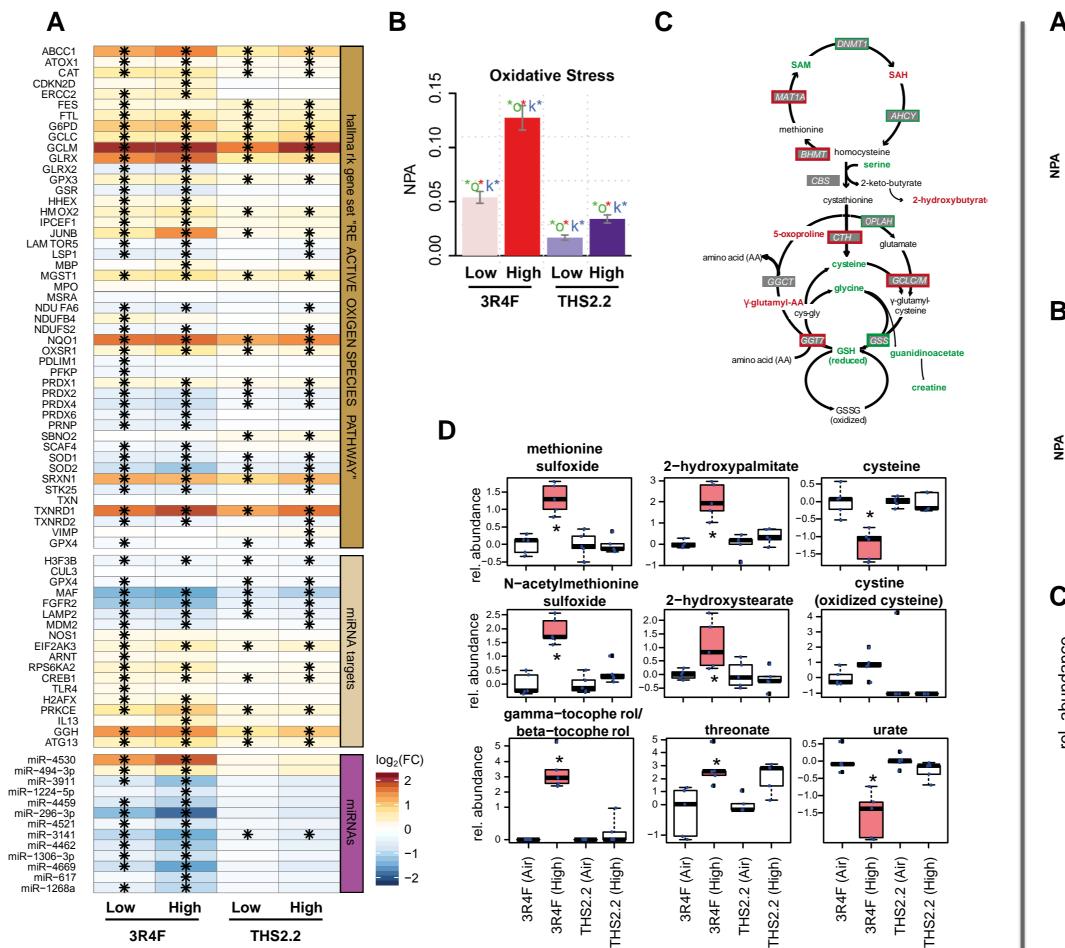
commercially marketed tobacco products". The Tobacco Heating System (THS) 2.2 is a candidate MRTP based on a heat-not-burn technology that uses a precisely controlled	 Pro-inflammatory mediators. Pro-inflammatory mediators were measured in the basolateral medium using a Luminex[®]-based technology (Luminex, Austin, TX, USA). Microarray data processing and analysis. 	_	Name reported	Smoke/aerosol concentration (%)	Nicotine concentration measured in PBS (mg/L; average ± SEM)	Nicotine dose deposited in 100 μL PBS (μg/insert/28 min; average ± SEM)	epithelial organotypic cultures were exposed for 3 consecutive days to 28 min 3R4F CS or THS2.2 aerosol at two matching concentrations. Before each exposure, basolateral medium was collected for different assays (AK and cytokine assays) and replaced with fresh medium; apical PBS was replaced before each exposure.
heating device into which a specially designed tobacco product is inserted and heated to	Transcriptomics data were analyzed in the context of	3R4F low	3R4F (Low)	25	49.4 ± 1.89	4.94 ± 0.189	Different endpoints were analyzed at the indicated time points during
generate an aerosol [2].	hierarchically structured network models as described in [3]. The effects of exposure were quantified by	concentration 3R4F high concentration	3R4F (High)	35	84.6 ± 1.43	8.46 ± 0.143	three experimental repetitions (with three independent exposure runs each, n=9 total). ($$) for 24 h mRNA/miRNA endpoint indicates that
Objectives. The objective of the study was to	scoring the impact on each subnetwork (referred to as	THS2.2 low	THS2.2 (Low)	75	54.6 ± 2.60	5.46 ± 0.260	only one experimental repetition (n=3) was performed (see [1] for
assess – using a systems toxicology approach— how aerosol from THS2.2, compared to	"network perturbation amplitude", NPA) [4] . Metabolomic analysis. Metabolites were analyzed in	concentration THS2.2 high concentration	THS2.2 (High)	100	100.4 ± 4.83	10.04 ± 0.483	24h results). 1, aerosol inlet; 2, culture well; 3, culture insert; 4, apical PBS; 5, organotypic culture; 6, membrane; 7, medium. (B) Selected
reference (3R4F) cigarette smoke (CS), affects human gingival epithelial organotypic cultures.	collaboration with Metabolon inc. (Durham, USA) [1].						CS/aerosol concentrations and matching delivered nicotine doses. (C) Vitrocell dilution and exposure system.







OXIDATIVE STRESS



INFLAMMATION

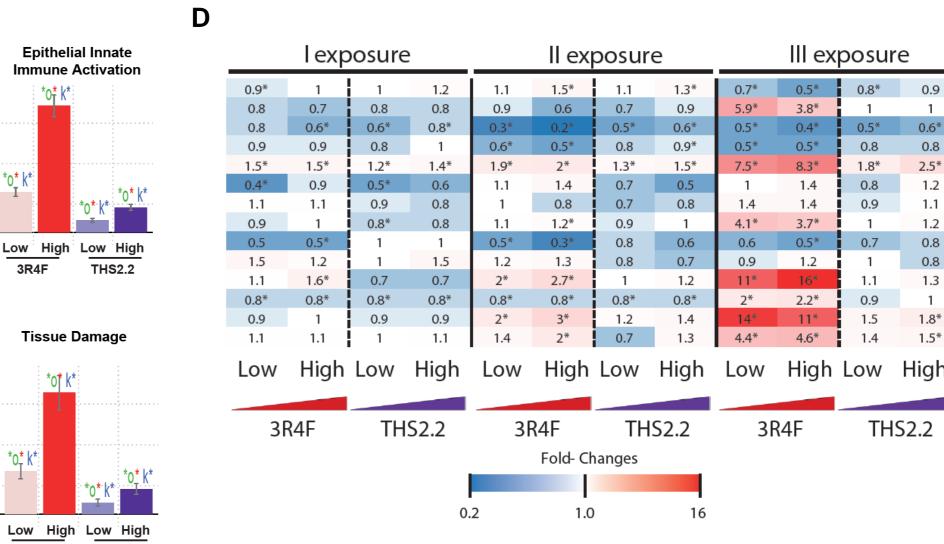


Figure 2. Cytotoxicity and tissue morphology of organotypic gingival cultures exposed to 3R4F CS and THS2.2 aerosol. Mean AK levels were assessed at 4 h (A) and 24 h (B) post exposure. AK levels were normalized to values of the positive control (Triton-X-treated cultures. considered 100% cytotoxicity). Error bars indicate SEM. * p<0.05, compared with the corresponding air control; # p<0.05, compared with matching concentrations of 3R4F CS. n=9. (C) Representative images of HE-stained gingival cultures. (D) Arrows indicate keratohyalin granules (3R4F (Low)) or parakeratosis (3R4F(High)). n=9. Abbreviations: M, membrane; SB, stratum basale, SS, stratum spinosum; SG, stratum granulosum; SC, stratum corneum.

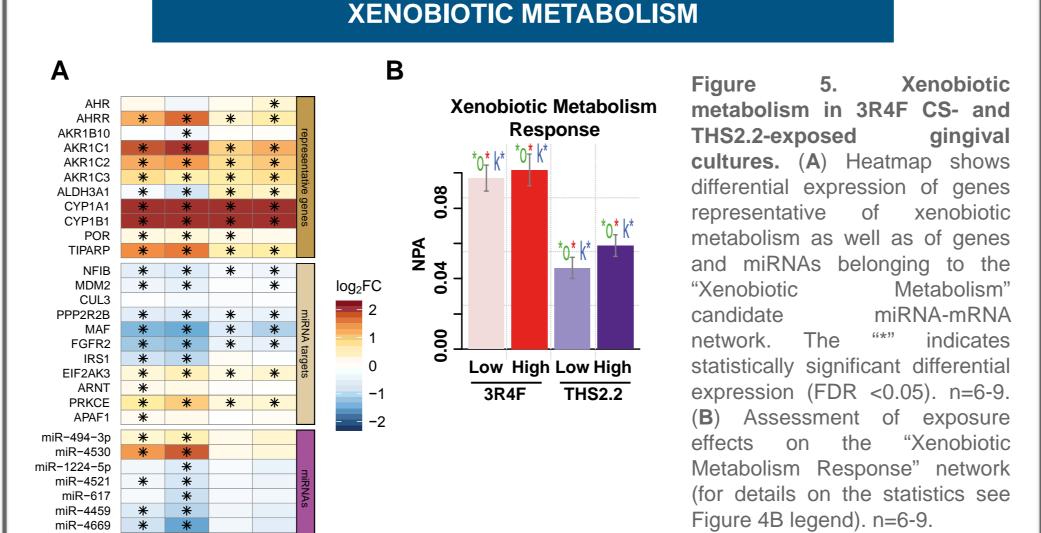
GENE EXPRESSION IMPACT OVERVIEW

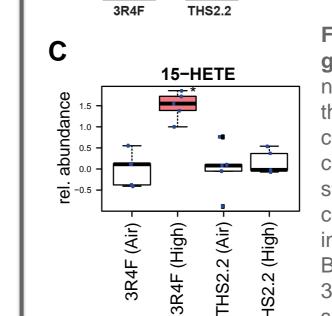
Biological Impact Factor (all 28 networks) Cell Fate (CFA - 5 networks) Cell Proliferation (CPR - 15 networks) Cellular Stress (CST - 6 networks) Inflammation (IPN - 2 networks) Apoptosis (CFA-1) Autophagy (CFA-2) Necroptosis (CFA-3) Response To DNA Damage (CFA-4) Senescence (CFA-5) Calcium (CPR-1) Cell Cycle (CPR-2) Cell Interaction (CPR-3) Clock (CPR-4 Causal network Epigenetics (CPR-5) Growth Factor (CPR-6) Hedgehog (CPR-7) perturbation Hox (CPR-8 Jak Stat (CPR-9) Mapk (CPR-10) MTor (CPR-11 Notch (CPR-12 Nuclear Receptors (CPR-13) PGE2 (CPR-14) Wnt (CPR-15) 0.6 Endoplasmic Reticulum Stress (CST-1 Hypoxic Stress (CST-2 0.4 NFE2L2 Signaling (CST-3 Osmotic Stress (CST-4) 0.2 Oxidative Stress (CST-5 Xenobiotic Metabolism Response (CST-6) 0.0 Epithelial Innate Immune Activation (IPN-1 Tissue Damage (IPN-2) KEGG collection - Q1, (FDR<0.05)*|stat| (209 pathways) KEGG collection - Q2. (FDR<0.05)*istati (209 pathways) Network-matching subset - Q1, (FDR<0.05)*|stat| (22 pathways) Network-matching subset - Q2, (FDR<0.05)*|stat| (22 pathways) mmune-related subset - Q1, (FDR<0.05)*|stat) (8 pathways) Gene-set nmune-related subset - Q2, (FDR<0.05)*|stat| (8 pathways) Metabolism subset - Q1. (FDR<0.05)*(stat) (44 pathways) etabolism subset - Q2, (FDR<0.05)*|stat| (44 pathways) analysis tabolism/Signaling subset - Q1, (FDR<0.05)*|stat) (7 pathways) tabolism/Signaling subset - Q2, (FDR<0.05)*|stat| (7 pathways) Signaling subset - Q1, (FDR<0.05)*|stat| (145 pathways Signaling subset – Q2, (FDR<0.05)*|stat| (145 pathways) ress Response subset – Q1, (FDR<0.05)*|stat| (5 pathways) Stress Response subset – Q2, (FDR<0.05)*|stat| (5 pathways) Differentially Expressed Genes - (FDR<0.1)*|FC| [maximum=11641] Differentially Expressed Genes - (FDR<0.05)*[FC] [maximum=10196] Differentially Expressed Genes - (FDR<0.01)*[FC] [maximum=7683] Differential Differentially Expressed Genes - (FDR<0.001)* FC [maxmum=5487] Differentially Expressed microRNAs - (FDR<0.1)*|FC| [maximum=185] expression Differentially Expressed microRNAs – (FDR<0.05)*[FC] [maximum=148] Differentially Expressed microRNAs – (FDR<0.01)*[FC] [maximum=104] Differentially Expressed microRNAs - (FDR<0.001)*[FC] [maximum=67] Low High Low High

Figure 3. Overview of the impact of 3R4F CS or THS2.2 aerosol exposures on differential expression of genes. Values are normalized to the interval [0, 1] in a row-wise manner. The uppermost panel displays the overall biological impact factor (BIF) and the relative BIFs for four network families (cell fate and angiogenesis (CFA), cell proliferation (CPR), cellular stress (CST), and pulmonary inflammation (IPN)) assessed by a causal network approach [4,5]. The network perturbation amplitudes (NPA) for the individual networks are shown in the next panel. The "*" indicates statistically significant network perturbations. Overall results of gene set analyses (GSA) are displayed in the next panel for the KEGG collection and the two standard statistical tests (Q1 and Q2). Also shown are specific subsets of the KEGG collection: first, the 22 pathways matching the mechanistic networks, and second, the five broad categories of the 228 pathways contained in the KEGG collection. Displayed values were defined as sums of absolute values of gene set-level statistics (i.e. fold-change mean) for the statistically significant gene sets in each category. The two lower panels show the number of differentially expressed genes (DE) and miRNAs (miRDE) for four distinct statistical significance thresholds, to identify possible threshold effects. Again, sums of absolute values of fold-changes of statistically significant genes or miRNAs are displayed. n=6-9.

3R4F THS2.2

Figure 4. Differential induction of oxidative stress by 3R4F CS and THS2.2 aerosol (A) Induction of oxidative stress response program: differential expression heatmap for genes of the reactive oxygen species pathway (HALLMARK_REACTIVE_OXIGEN_SPECIES_PATHWAY; M5938), as well as for genes and miRNAs belonging to the "Oxidative Stress" candidate miRNA-mRNA network (see [1] for details). The "*" indicates statistically significant differential expression (FDR <0.05). n=6-9. (B) Assessment of exposure effects on the "Oxidative Stress" network. Bars show overall network perturbation amplitudes (NPA scores) based on transcriptomics data. Error bars delimit their 95% confidence intervals. Three statistics are shown: the red star indicates statistical significance with respect to the biological replication (i.e. 95% confidence intervals do not contain the 0 value), while the green and red stars indicate significant specificity statistics with respect to the network structure ("O" and "K" statistics). n=6-9. (C) Summary of exposure effects on glutathione and related metabolic reactions (see [1] for details). Significantly up- or down-regulated metabolites and genes are marked with red and green, respectively. (D) Metabolomics profiling was conducted 4 h after exposure of the tissue to high 3R4F CS and THS2.2 aerosol concentrations. Boxplots summarize the response of metabolites sensitive to oxidative stress (blue dots indicate individual samples, n=5). Significant differences between exposed groups and their respective sham groups are indicated by filled colored boxes and a star ("*" means FDR < 0.05).





3R4F

Low

Figure 6. Profile of inflammation in 3R4F CS- and THS2.2 aerosol-exposed **gingival cultures.** (A–B) Assessment of exposure effects on the inflammation networks "Epithelial Innate Immune Activation" and "Tissue Damage" (for details on the statistics see Figure 4B legend). n=6-9. (C) Metabolomics profiling was conducted 4 h after exposure of the tissue to high 3R4F CS and THS2.2 aerosol concentrations. Boxplot summarizes the response of 15-HETE (For details on the statistics see Figure 4D legend). (D) Heatmap showing fold-changes of mean concentrations of proinflammatory mediators in exposed cultures relative to those in their corresponding air controls 24 h after each exposure (I, II, III exposure). Blue and red colors indicate negative or positive fold-changes, respectively, in 3R4F CS- and THS2.2 aerosol-exposed samples compared with air-exposed samples. n=9.

TNFA

CCL2

IP-10

IL8

IL1A

CXCL

CSF2

CSF3

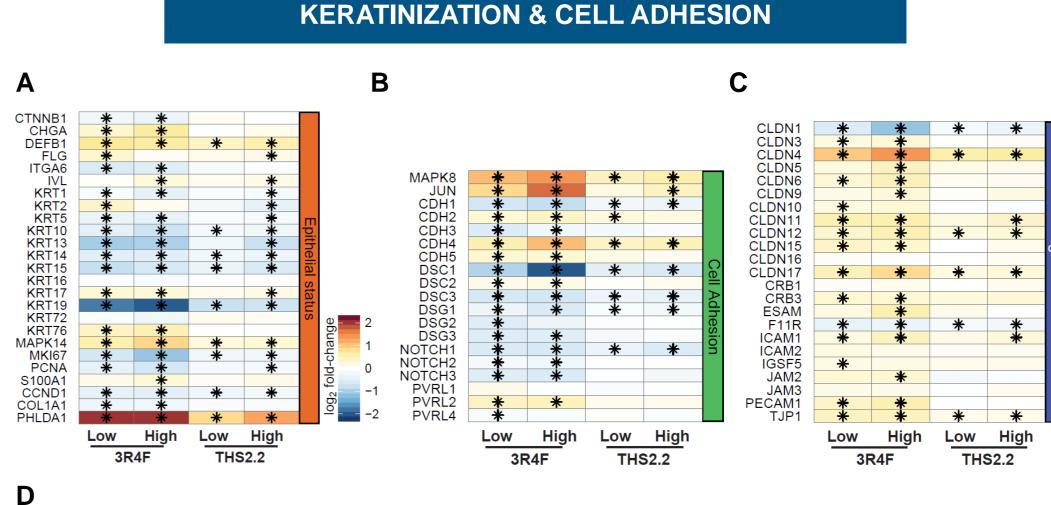
1.2

1.2

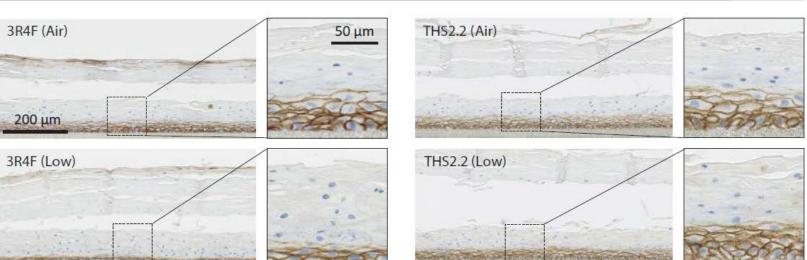
1.3

1.8*

1.5*



24 h Post-Exposure



	3R	4F	тн	S2 2	
	Low	High	Low	High	
miR-296-3p	*	*			
miR-4669	*	*			
miR-4459	*	*			S
miR-617		*			miRNAs
miR-4521	*	*			niŖ
R-1224-5p	•	*			_
miR-4530	*	*			
niR-494-3p	*	*			
APAF1	*				
PRKCE	*	*	*	*	
ARNT	*				1
EIF2AK3	*	*	*	*	targets
IRS1	*	*			arg
FGFR2	*	*	*	*	A
MAF	*	*	*	*	miRNA
PPP2R2B	*	*	*	*	з
CUL3					

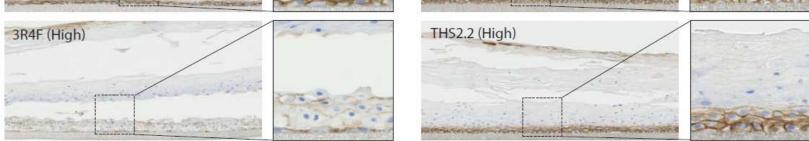


Figure 7. Keratinization and cell-cell adhesion in 3R4F CS- and THS2.2 aerosol-exposed samples. (A, B, C) Heatmaps showing differential expression of genes related to epithelial status/cell type, cell adhesion, or tight junctions. (for details on the statistics see Figure 4B legend). N=6-9. (D) Representative images of Ecadherin-stained gingival culture sections observed 24 h after the last exposure. Magnification is set at 20× and $63 \times$ for the insets. n=9.

CONCLUSIONS

REFERENCES

• Systems toxicology approach was applied for the assessment of THS2.2 aerosol compared to CS on an organotypic gingival epithelium model. Multiple endpoints (e.g., cytotoxicity, transcriptomics, and metabolomics) were combined toward a comprehensive assessment of the exposure effects.

• Major morphological alterations (loss of cell adhesion, keratinization, Figure 7) and cytotoxicity (max. ~30%, Figure 2) were observed after CS exposure but were limited, if none, upon exposure to THS2.2 aerosol.

• Transcriptomic and metabolomic analysis indicated a general reduction of the impact in THS2.2 aerosol-exposed samples with respect to CS (~79% lower biological impact for the high THS2.2 aerosol concentration compared to CS, and 13 metabolites significantly perturbed upon THS2.2 aerosol exposure vs. 181 for 3R4F CS).

• Proinflammatory mediator analysis showed a higher impact in CS-exposed cultures compared to THS2.2 aerosol, with 11 analytes significantly altered by CS vs. 5 (common to CS-exposure condition) by THS2.2 aerosol, showing a reduced fold-change with respect to CS (Figure 6).

 Biological effects induced by CS, such as oxidative stress, xenobiotic metabolism, and inflammation-related processes, are relevant to the pathophysiology of periodontal diseases.

• Overall, THS2.2 aerosol had a statistically significantly lower impact on molecular processes associated with the pathophysiology of human gingival organotypic cultures compared to CS.

[1] Zanetti F, Titz B, Sewer A, Lo Sasso G, Scotti E, Schlage WK, Mathis C, Leroy P, Majeed S, Torres LO, Keppler BR, Ashraf E Trivedi K, Guedj E, Martin F, Frentzel S, Ivanov NV, Peitsch MC, Hoeng J. Comparative systems toxicology analysis of cigarette smoke and aerosol from a candidate modified risk tobacco product in organotypic human gingival epithelial cultures: A 3-day repeated exposure study. Food Chem Toxicol. 2016 Dec 23. pii: S0278-6915(16)30488-4. doi: 10.1016/j.fct.2016.12.027. [2] Smith MR, Clark B, Lüdicke F, Schaller JP, Vanscheeuwijck P, Hoeng J, Peitsch MC. Evaluation of the Tobacco Heating System 2.2. Part 1: Description of the system and the scientific assessment program. Regul Toxicol Pharmacol. 2016 Nov 30;81 Suppl 2:S17-S26. doi: 10.1016/j.yrtph.2016.07.006. [3] Hoeng J, Deehan R, Pratt D, Martin F, Sewer A, Thomson TM, Drubin DA, Waters CA, de Graaf D, Peitsch MC. A networkbased approach to quantifying the impact of biologically active substances. Drug Discovery Today. 2012 May 17; 413-8. doi: 10.1016/j.drudis.2011.11.008. [4] Martin F, Sewer A, Talikka M, Xiang Y, Hoeng J, Peitsch MC. Quantification of biological network perturbations for mechanistic insight and diagnostics using two-layer causal models. BMC Bioinformatics. 2014 Jul 11;15:238. doi: 10.1186/1471-2105-15-238. [5] Martin F, Thomson TM, Sewer A, Drubin DA, Mathis C, Weisensee D, Pratt D, Hoeng J, Peitsch MC. Assessment of network perturbation amplitudes by applying high-throughput data to causal biological networks. BMC Syst Biol. 2012 May 31;6:54. doi: 10.1186/1752-0509-6-54.

The work reported in this publication involved a candidate Modified Risk Tobacco Product developed by Philip Morris International (PMI) and was solely funded by PMI. All authors are employees of, or (W. K. Schlage) contracted and paid by Philip Morris International., except Brian R. Keppler (Metabolon Inc.).