

Untargeted Metabolomics Application to Monitor Urine Samples by Ultra-High-Pressure Liquid Chromatography **Coupled to High-Resolution Mass Spectrometry**

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Objectives

- In order to meet the demands of high-throughput metabolomics, we established an efficient, easily transferable LC-HR-MS method for urine sample analysis.
- We employed the latest model of Vanquish tandem LC system (ThermoFisher) with considerable time saving benefit.
- Retention time (RT) drift across the columns was accounted for and RTs were re-aligned using dynamic time warping (DTW) [1,2].
- In order to improve the normalization strategy and implicitly obtain a more robust method for metabolite identification we have assessed the benefits of spiking urine samples with ¹³C yeast extract (TruQuant IQQ Workflow kit, IROA) [3].
- Urine metabolite concentrations vary widely (up to 15-fold) due to water uptake [4], as part of our normalization strategy, here we asses the importance of urine volume normalization based on creatinine levels using matrix matched calibration curve approach [5].

IROA ¹³C yeast extract application for untargeted urine metabolomics

Here we explore the possibilities offered by spiking pooled urine sample with a complex internal standard (IS) obtained from metabolically labeled yeast extract that contains hundreds of ¹³C metabolites (IROA).



Methods

- Pooled human urine sample was thawed and centrifuged at 15,000 g for 15 minutes at 4 °C.
- Separation of metabolites was performed using two Hypersil Gold C18 columns (150 x 2.1 mm, 1.9 µm) operating in parallel and running with a fast linear gradient of acetonitrile containing 0.1% formic acid ramping from 5% to 95% in 15 min, alternating between the two columns at 0.5 ml/min. Column was kept at 50 °C.
- Dual-column operation improves the chromatography repeatability, maintains full occupancy of the MS instrument, enabling approx. 100 injections per day.
- Same urine sample (2 µl) was injected 80 times on each column in positive, and repeated in negative ESI mode.



• Mass spectrometric detection was done on a Q Exactive HF mass spectrometer operating in positive and negative electrospray ionization FS acquisition modes with a mass range m/z 70-1050 and resolution 120k FWHM @ 200 m/z. Presented data is in positive mode unless otherwise stated.

Results

Retention time re-alignment across different columns or systems

• Dynamic time warping (DTW) is an algorithm for measuring similarity between two temporal

- Figure 4: Left: example of ¹²C and ¹³C labelled Leucine (Leu) and Isoleucine (IIe) detection principle with the IROA, right: ClusterFinder analysis software screenshot.
- In order to build a calibration curve, 0 to 100 µl volume of urine were dried down under nitrogen flow and re-suspended in 40 µl of ¹³C labelled metabolite solution as per fabricant recommendation [3].



Figure 5: A: Leucine (Leu), Isoleucine (IIe) chromatographic separation; B: Leu and IIe peak areas determined by red dots, Secondary axis (right) and yellow dots represent areas corresponding to ions of the ¹³C internal standards. C: Leu and Ile normalized calibration curves. Linear regression R^2 values presented on graphs.

Urine sample volume normalization

• Volume normalization consideration takes into account detected number of features as a function of measured creatinine levels, as well as ion suppression observed in ESI-MS



sequences. In chromatography, DTW is using the Euclidean distance to construct the shortest path between a reference chromatogram and a target chromatogram that minimizes the sum of absolute differences. DTW aligns each point of the query chromatogram.





Figure 6: Number of total features detected (blue bars) by Compound Discoverer 3.0 (ThermoFisher) for increasing creatinine concentration levels. Values in table on the right show % gain in number of features compared to the previous creatinine concentration level.

• Ion suppression is a poorly understood phenomena consisting of the loss of ionization of analytes. Stable isotope labeled IS spiking is considered to be one of the best approaches to compensate for ion suppression. Below we look at the percent ion suppression of ¹³C labelled Tryptophan



Figure 7: A. Percent ion suppression of ¹³C Tryptophan IS in positive (red) and negative (blue) ionization modes. B. Global effect of ion suppression across the analysis with TIC insert overlap.

Conclusions

- \checkmark We doubled the throughput due to dual column switching operating mode (~100 runs/day).
- DTW enables accurate peak alignment mandatory for metabolomics applications.





Figure 3: Inter-sample correlation of total ion current (TIC), unaligned (blue), aligned with Fourier Transform (FT) (red) and aligned with DTW (green)

.6min)

YES 0,04

A.	m/z166.08626		m/z120.08130				_			DTW ali	ignment	
	Phe_quan		Phe_qual		Ratio			Β.	Avg. Area		RT (7	
	Area	Height	Area	Height	Area	Height			NO	YES	NO	
CV (%)	9,9	9,7	8,8	8,6	3,7	4,9		CV (%)	7,7	2,85	0,21	

Table 1: A:Coefficient of variation (CV) of peak area and height of Phenylalanine qualifier and quantifier ions within 200 injections on both CoIA and CoIB; B: Average area CV% entire chromatogram and RT CV% at 7,6 min before and after DTW, 200 injections.

- ✓ Average CV of the area of all features was 2.85% from 200 injections of the same urine sample.
- ✓ ¹³C IROA yeast extract proves to be beneficial for:
 - easier metabolite identification through better annotation (ClusterFinder software)
 - ion suppression monitoring and correction along the whole chromatogram
 - accurate normalization from associated ¹³C proper IS or closely eluting alternative ¹³C compounds calibration curve building
- Indicative information was obtained regarding optimal urine creatinine concentration for sample volume normalization purposes.

References

[1] Clifford D. et al. 2009 Anal. Chem., 81, 3, 1000-1007 [2] Jiang W. et al. 2013 Chromatographia 76 1067–1078 [3] Beecher C. and de Jong FA. **2019** *Methods Mol Biol.* <u>1996</u> 17-28 [4] Warrack B. et al. 2009 J. Chromatogr. B, <u>877</u>, 547–552 [5] Kennedy A.D. et al. 2016 Gen. Test and Mol. Biomarkers. 20, 9, 485-495.

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