Exposure to Cigarette Smoke and Aerosol from the Candidate Modified Risk Tobacco Product THS2.2 on A/J Mice in a Life-Time Inhalation Study

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Poster Board P-04-05-08 Abstract ID 390

Introduction

Smoking causes many serious diseases including lung cancer and chronic obstructive pulmonary disease (COPD). Mouse models have been employed to study the carcinogenicity of chemical compounds and infer underlying mechanisms of lung tumor development in humans¹. Different strains of mice display markedly varied sensitivity to lung tumor development. For example, mice of the C57Bl/6 strain are rather resistant to tumor induction, while Balb/c mice are considered intermediate in susceptibility². In contrast, the A/J mouse is highly susceptible to lung tumor induction and has been widely used as a screening system in carcinogenicity testing. Although this mouse strain is developing lung tumors spontaneously, an 18-month inhalation period was previously found to be sufficient in eliciting a CS concentration-dependent lung tumor response³. Moreover, gene and microRNA (miRNA) expression analysis demonstrated that lung tumors developing in MS-exposed A/J mice exhibit marked differences compared to spontaneously arising tumors, and that these differences can be harnessed in the form of a gene signature to potentially predict the extent of exposure.⁴ Here, we report on an 18-month in vivo inhalation study, performed on A/J mice, to assess the chronic toxicity of exposure to aerosol from THS2.2, a candidate modified risk tobacco product (cMRTP), and to compare it with the toxicity of smoke from the 3R4F reference cigarette (CS). This mouse model has previously been shown to be susceptible to develop lung tumors and has been used in many carcinogenicity studies.

General Health, Survival, Hematology and Clinical Chemistry



Figure 4. Body weight development throughout the course of the study.

Figure 5. Survival statistics for (A) female and (B) male A/J mice. Data are presented as Kaplan-Meier survival probablility estimates throughout the course of the study. + indicates censored data (technical deaths).



Study Design and Endpoints

Female A/J mice were whole-body exposed to filtered air (Sham), to three concentrations of THS2.2 aerosol (6.7, 13.4, 26.8 µg/L nicotine) and one concentration of 3R4F CS (13.4 µg/L

nicotine) for 6 h/day, 5 d/week, for up to 18 months; male mice were exposed to filtered air Figure 6. (A) Red blood cell (erythrocyte), (B) total white blood cell, (C) absolute lymphocyte and (D) absolute neutrophil counts in blood of in (Sham) and the highest concentration of THS2.2 aerosol (Figure 1). Care and use of the mice A/J mice following 15 (male mice) or 18 (female mice) months of exposure. Data are presented as median±mean absolute deviation (MAD). Lines indicate was in accordance with the National Advisory Committee for Laboratory Animal Research statistical differences between groups (p<0.05, t-test, not adjusted).

Guideline 2004⁵. All animal experiments were approved by the Institutional Animal Care and Use Committee (IACUC). Animals were observed on a daily basis, body weight progression was A »Liver Function monitored once per week, exposure parameters were measured at regular intervals. Toxicological assessment was performed according to published standard methods^{6, 7} including histopathological evaluation, which is currently still ongoing. Interim dissections were performed after 1, 5 and 10 months of exposure. At months 1 and 5, animals were also allocated for the following endpoints: bronchoalveolar lavage fluid (BALF) analysis by FACS and multi-analyte (cytokines/chemokines) profiling; lung function tests. At each time point, nasal epithelium, trachea, lungs and blood were collected from a subset of animals for an extensive molecular high-throughput analysis (transcriptomics, proteomics).





Figure 7. (A) Total protein, (B) alkaline phosphatase (ALP), (C) glucose and (D) total cholesterol levels in serum of in A/J mice following 15 (male mice) or 18 (female mice) months of exposure. Data are presented as mean±SEM or median±MAD. Lines indicate statistical differences between groups (t-test, *p*<0.05, not adjusted). Diamonds indicate significant differences between male and female mice (t-test, *p*<0.05, not adjusted).









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Figure 1. Schematic overview of study design, endpoints and interim dissection time points.

Aerosol Exposure and Uptake



Figure 8. A/J mouse nasal epithelial responses. (A) Histopathological findings, (B) systems response profiles, (C) biological impact, (D) mechanistic responses and (E) differential protein abundances following 15 (male mice) or **18 (female mice) months of exposure.** Shown are mean severity scores ±SEM for nasal respiratory epithelial hyperplasia and



vacuolization (level 1) of exposed females at month 10 (A). Volcano plots were generated to demonstrate systems responses in the nasal epithelium (B). Each dot in a plot represents a gene. Gene expression changes (x-axis) are expressed as [log_(fold-change)]; increased when >0 and decreased when <0 as compared with Sham. The statistical significance (y-axis) is represented as [-log₁₀(adj.p-value FDR)]. The horizontal line represents statistical significance [-log₁₀(0.05)]. The bar plot (C) depicts BIF values relative to the maximum response (RBIF = 100%, REF). The δ values (-1 to 1) indicate how similar the underlying network perturbations are with respect to the *REF. Scores were computed using transcriptomics profiling data from nasal epithelia. (D) Starplots illustrating the response of the mechanisms, for each the treatment* groups (the slices). The surface area of each slice is proportional to the contribution of that mechnism to the overall RBIF of the treatment. The heatmap (E) depicts fold changes in protein abundances (protein symbols listed on the right) relative to corresponding air-exposed controls expressed as log2[fold change (value; fc)]. Increased and decreased fold changes for the top 50 proteins are represented by orange and blue shades, respectively, with darker colors indicating greater absolute fold changes.







tracheal Figure mouse epithelial (A) responses. Histopathological findings at month 10, (B) systems response profiles, (C) biological impact and (D) mechanistic response following 15 (male mice) or 18 (female mice) months of exposure.

Figure 3. (A) Total urinary nicotine metabolites following 13 (male mice) or 15 months (female mice) of exposure. (B) Blood carboxyhemoglobin levels following 16 months (female mice) of *exposure.* Data are presented as mean ± MAD. Lines indicate statistical group comparisons (t-test) where *p*-values fell below 0.05 (not adjusted).

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EUROTOX 2017 10-13 September 2017 Bratislava, Slovakia

Conclusions

The characterization of the aerosol components as well as biomarkers of aerosol uptake indicated that the delivery and uptake of 3R4F CS and THS2.2 aerosol by the mice were in line with expectations. The test atmosphere was well tolerated as reflected by the progressive weight gain over time. Exposure to CS induced changes in red and white blood cell profiles, liver function and metabolic parameters, but only minimal changes were observed following THS2.2 aerosol exposure. Exposure to CS also resulted in histopathological changes in the nose and trachea, while no significant effects were observed following THS2.2 aerosol exposure. Significant changes in the nasal and tracheal epithelial transcriptomes and the nasal epithelial proteome were observed following CS exposure. The perturbed mechanisms are consistent with CS exposure, but only minor perturbations were noted in nose and trachea following THS2.2 aerosol exposure. With the histopathological evaluation of the lungs and other organs still ongoing, no final conclusion regarding the chronic toxicity and carcinogenicity of THS2.2 can be drawn at this point.

> **Competing Financial Interests** The research described in this poster was sponsored by Philip Morris International