

SCIENCE

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Introduction and Objectives

With increasing popularity of electronic cigarettes (ECs), it is important to assess the potential toxicity of EC aerosol exposure. In the context of a harm reduction approach, we examined the effects of whole EC aerosol exposure relative to the effects of mainstream cigarette smoke (CS) exposure using human organotypic small airway cultures. Cultures were exposed at the air-liquid interface to 112 puffs of either undiluted aerosols generated from a MarkTen[®] cartridge containing various e-liquids (with aerosol formers alone ["Carrier"], with 4% nicotine ["Base"], with 4% nicotine and flavors ["Test Red"] or to diluted CS, in Vitrocell[®] exposure systems. We conducted a series of independent exposure repetitions to strengthen the accuracy of the observation. Concentrations of the deposited nicotine and carbonyls in the exposure chamber were measured as markers of exposure. Biological endpoints investigated included histology, cytotoxicity, inflammatory mediators, and gene microarray. Alterations in morphology or cytotoxicity were not observed in small airway cultures exposed to undiluted EC aerosols despite resulting in higher nicotine deposition in the chamber than that found following CS exposure. Increased loss of cilia was recorded in cultures exposed to CS but not in cultures exposed to any EC aerosol. Media of cultures exposed to CS had generally greater levels of inflammatory mediators in comparison to cultures exposed to EC aerosols. CS exposure also elicited a greater number of differentially expressed genes. Based on a network-based enrichment analysis, the transcriptome data showed that the exposures impacted different cellular processes (e.g., cell fate, proliferation, stress, and inflammatory response) with greater impacts following CS exposure than following EC aerosol exposure. These collective endpoints demonstrated that EC aerosols had significantly lower biological impact in small airway cultures, in comparison to CS exposure.

> **Culture Morphology Following Exposure** 100 µm **Apoptosis Score** AT 1- CONTROL STRATION STRATE LAS 80% 60% 40% 20% 100 µm 100 µm Histopathology Score 0

for 28 min (112 puffs) (12 µg nicotine/mL PBS)

13% 3R4F CS

100% Test Red EC aerosol for 28 min (112 puffs) (134 µg nicotine/mL PBS)

100% Base EC aerosol for 28 min (112 puffs) (104 µg nicotine/mL PBS)

100% Carrier EC aerosol for 28 min (112 puffs) (0 µg nicotine/mL PBS)

100% Air for 28 min (112 puffs) (Control)

Representative images of the H&E-and Alcian blue-stained sections are shown. In small airway cultures, thinning of the epithelium was seen in following exposure to the 28-minute (112-puff) 13% 3R4F CS, corresponding to around 12 µg/mL deposited nicotine. In contrast, exposure to undiluted Test Red or Base aerosols did not alter the culture morphology despite delivering higher nicotine concentrations.

Increased apoptosis was detected following exposure to 7% or 13% 3R4F CS for 28 min (112 puffs) compared with the exposure to air. Conversely, it was not detected following exposure to 100% Test Red, Base, or Carrier aerosols for the same duration.

In conclusion, when compared to air exposure, exposure to Test Red, Base, or Carrier EC aerosols resulted in a lower impact than exposure to CS on in vitro human organotypic small airway epithelial cultures. Overall, marked differences in the measured endpoints (culture morphology, ciliary beating frequencies, secretion of inflammatory mediators, and global gene expression changes) were not observed following exposure to Test Red, Base, or Carrier EC aerosols. The observation further suggested minimal specific effects of nicotine or flavor ingredients in the alterations of various biological endpoints measured in the study.

Comparison of the impacts of an acute exposure to electronic cigarette aerosol and cigarette smoke on small airway epithelial cultures: In vitro systems toxicology assessment





Results

Reduced ciliary beating frequencies were detected following exposure to 7% 3R4F CS for 28 minutes (112 puffs) compared with exposure to air. The reduced frequencies remained detected at 24 hours post-exposure and finally recovered at 48 hours post-exposure. Exposure to 100% Test Red, Base, or Carrier aerosols, as well as the air-exposed controls, for the same duration resulted in a slight reduction in ciliary beating frequencies; however, this was not statistically significant and recovered at the 24 hours post-exposure time point.

Conclusions

robustness. For the histological analysis, the cross-sections of the organotypic epithelium cultures were analyzed after hematoxylin and eosin (H&E) and Alcian blue staining. Ciliary beating measurement was conducted using the Sisson Ammons Video Analysis system on a total of 512 video frames recorded from the center of the insert surface. Concentrations of inflammatory mediators were measured from the basolateral medium of the exposed cultures using Luminex[®] xMAP[®] technology and commercially available assay panels (EMD Millipore Corp) according to the manufacturer's instructions. Messenger RNA microarrays were done using 100 ng of total RNA (per sample) that were reverse-transcribed and amplified to cRNA using the Affymetrix® HT 3' IVT PLUS kit.



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	Time Point of Measurement				
PBS samples	Pre-Exposure	Post-Exposure			
	-	0 h	2 h	24 h	48 h
<mark>osure chamber</mark>	-	\checkmark	-	-	_
lial cultures	_	_	_	_	\checkmark
	\checkmark	\checkmark	-	\checkmark	\checkmark
	-	-	\checkmark	\checkmark	_
ateral medium	samples				
torc	_				/

Secretion of Inflammatory Mediators



Fold changes of the concentrations of secreted inflammatory mediators are shown. Exposure to 7% 3R4F CS for 28 minutes (112 puffs) markedly increased the concentrations of the mediators in the basolateral medium compared with the levels following exposure to air. Exposure to Test Red, Base, or Carrier aerosols generally elicited smaller changes in the mediator concentrations compared with the changes following exposure to 7% 3R4F CS for the same duration.

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