Toxicological evaluation of the Tobacco Heating System 2.2, a candidate modified risk tobacco product in a OECD inhalation study complemented with systems toxicology

U. Kogel¹, E.T. Wong², B. Titz¹, S. Boue¹, A. Elamin¹, G. Vuillaume¹, P. Leroy¹, N. V. Ivanov¹, P. Vanscheeuwijck¹, J. Hoeng¹, M.C. Peitsch¹

¹Philip Morris International R&D, Philip Morris Products S.A., Quai Jeanrenaud 5, 2000 Neuchâtel, Switzerland ²Philip Morris International Research Laboratories Pte Ltd, 50 Science Park Road, Science Park II, Singapore 117406

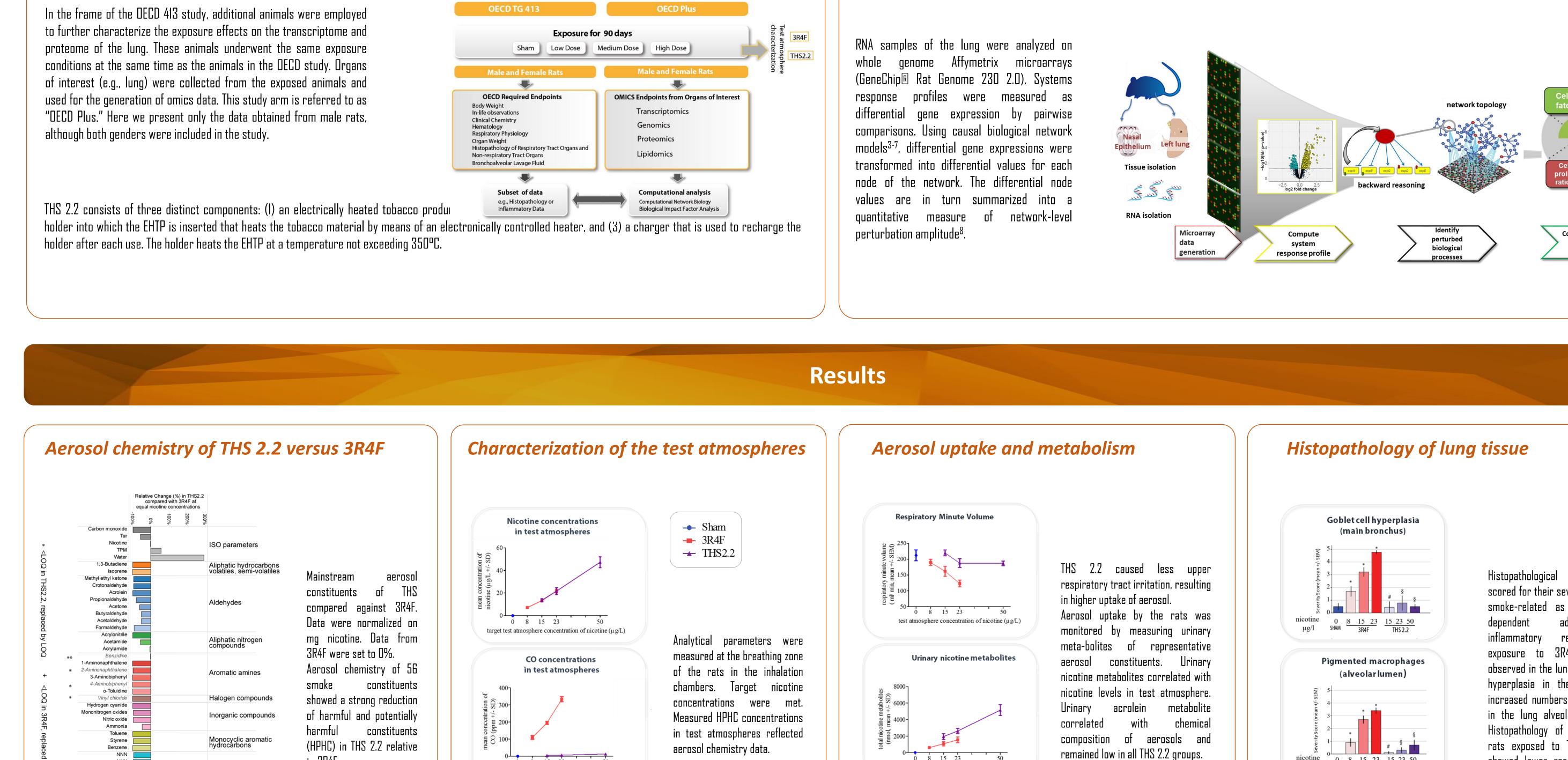
Introduction and study design

Methods

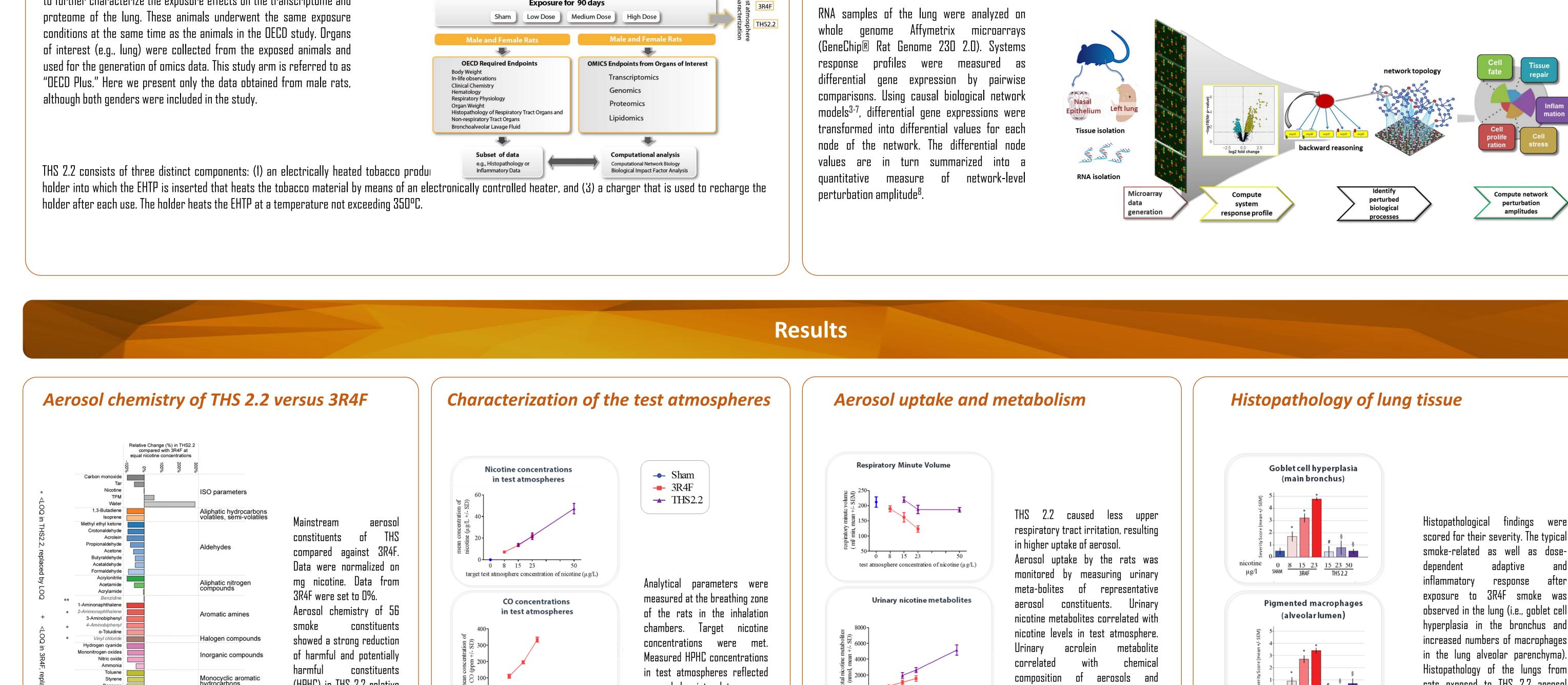
Smoking causes serious diseases, such as lung cancer, cardiovascular disease, and chronic obstructive pulmonary disease. Undoubtedly, the best way for smokers to prevent the adverse health effects of tobacco is to quit smoking. In recent years, tobacco harm reduction has also emerged as a policy that can complement traditional tobacco control interventions, such as prevention and cessation.

Here we show results of a 90-day rat inhalation study that was conducted in accordance with Organization for Economic Co-operation and Development (DECD) test guideline 4131 to characterize potential adverse effects caused by subchronic exposure to aerosol from the Tobacco Heating System (THS) 2.2, a heat-not-burn tobacco product, and to compare with those induced by the smoke generated from the 3R4F reference cigarette.

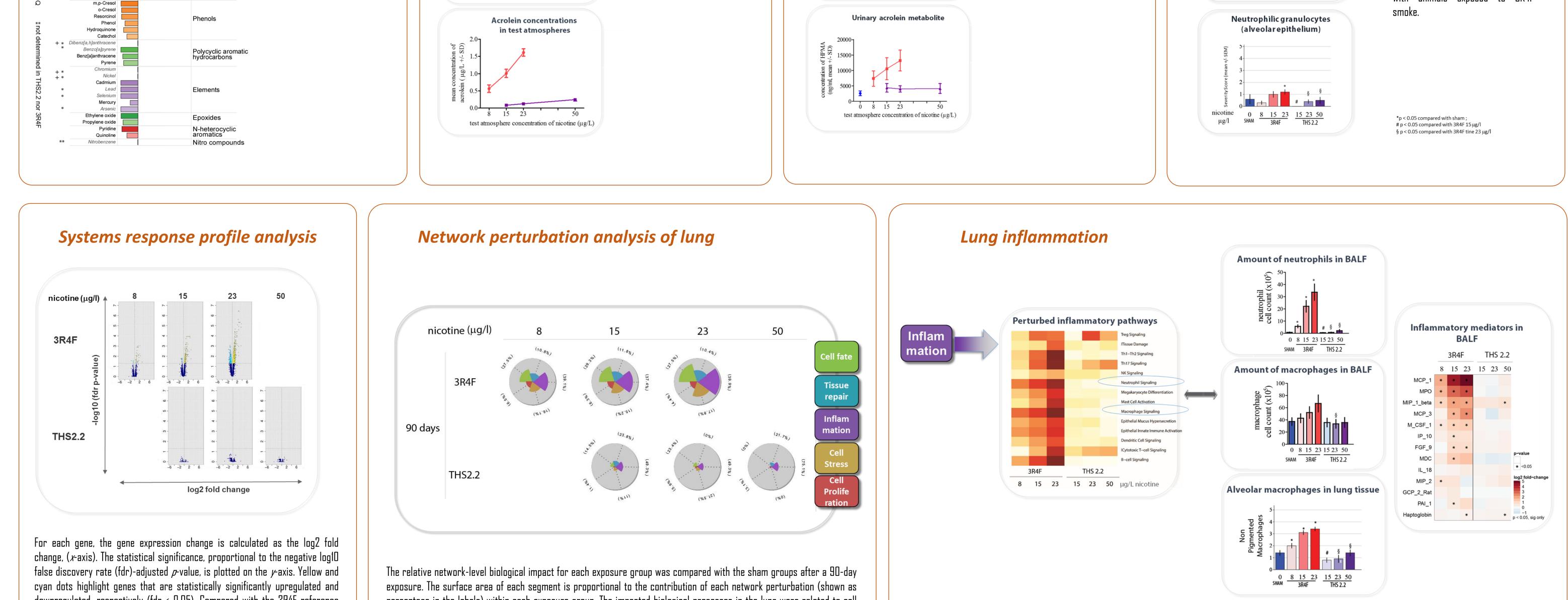
In the frame of the OECD 413 study, additional animals were employed to further characterize the exposure effects on the transcriptome and proteome of the lung. These animals underwent the same exposure conditions at the same time as the animals in the OECD study. Organs of interest (e.g., lung) were collected from the exposed animals and used for the generation of omics data. This study arm is referred to as "DECD Plus." Here we present only the data obtained from male rats,



Sprague Dawley rats were exposed for a period of 13 weeks to filtered air (sham), to three concentrations of mainstream 3R4F smoke (8, 15, 23 µg/L nicotine), or to THS 2.2 aerosol (15, 23, 50 µg/L nicotine). Care and use of the rats was in accordance with the National Advisory Committee for Laboratory Animal Research Guideline 2004². Exposure was confirmed by numerous biomarkers, well reflecting test atmosphere constituent concentrations. All animal experiments were approved by the Institutional Animal Care and Use Committee. Nicotine metabolites were determined by high-performance liquid chromatography after derivatization in 24-hour urine. Respiratory minute volume was determined using head-out plethysmography (EMKA Technologies, France). The histopathological evaluation was performed at defined anatomical sites of the left lung, according to a defined grading system. Free lung cells were determined in bronchoalveolar lavage fluid (BALF) by flow cytometry, and inflammatory mediators were measured by multianalyte profiling (RodentMAP® v3.0).



scored for their severity. The typical smoke-related as well as doseand after exposure to 3R4F smoke was observed in the lung (i.e., goblet cell hyperplasia in the bronchus and increased numbers of macrophages in the lung alveolar parenchyma). Histopathology of the lungs from rats exposed to THS 2.2 aerosol showed lower response compared with animals exposed to 3R4F



0 8 15 23

test atmosphere concentration of nicotine (µg/L)

downregulated, respectively (fdr < 0.05). Compared with the 3R4F reference cigarette smoke, THS 2.2 aerosol induced a minor response in lung tissue (although applying a higher dose of nicotine).

NNN

NNK

NAT

to 3R4F

N-nitrosamines

0 8 15 23

test atmosphere concentration of nicotine (μ g/L)

percentage in the labels) within each exposure group. The impacted biological processes in the lung were related to cell fate, cell stress, and inflammation after exposure to 3R4F reference cigarette smoke. Groups exposed to THS 2.2 aerosol showed a much smaller network perturbation compared with 3R4F-exposed groups.

Heatmap summarizing the inflammation subnetworks that were significantly perturbed in at least one treatment after nose-only exposure. Among those subnetworks, neutrophil and macrophage signaling were majorly impacted after exposure to the 3R4F reference cigarette but much less impacted after THS 2.2 exposure. This was also reflected in the differential cell count of BALF neutrophils and macrophages as well as in the amount of inflammatory mediators measured in the BALF.

nicotine

μg/l

SHAM

0 8 15 23 15 23 50

Conclusions	References
Histopathological evaluation showed usual dose-dependent adaptive and inflammatory responses for exposure to 3R4F reference cigarette smoke in rat lungs (i.e., goblet cell hyperplasia in the bronchi and increased amount of macrophages in the parenchyma). These findings were also consistent with previous inhalation studies ⁸⁻¹² . Lower severity scores were observed in the THS 2.2 aerosol-exposed animals, even at double the aerosol concentration on nicotine basis, compared with 3R4F smoke-exposed animals. Systems toxicology evaluation exemplified how classical toxicology endpoints (e.g., histopathology) can be complemented effectively by molecular measurements and computational analyses within a systems toxicology framework. Exposure of rats to aerosol from THS 2.2 resulted in a lower transcriptional systems response and lower biological network perturbations of lung tissue compared with exposure to 3R4F cigarette smoke. Computational network analysis also revealed that inflammation-related processes were less impacted after THS 2.2 aerosol exposed animals. Diverall, toxicology data indicated that THS 2.2 aerosol caused reduced lung toxicity in comparison with 3R4F cigarette smoke.	 1 DECD (2009) Test No. 413: Subchronic Inhalation Toxicity: 90-day Study. DECD Guidelines for the Testing of Chemicals, Section 4, DECD Publishing, Paris. 2 NACLAR Guidelines on the Care and Use of Animals for Scientific Purposes. Singapore: 2004. [accessed July 28, 2009]. 3 Westra, J.W. et al. (2011) Construction of a computable cell proliferation network focused on non-diseased lung cells, BMC Syst Biol. 5: 105. 4 Schlage, W.K. et al. (2013) A modular cell-type focused inflammatory process network model for non-diseased pulmonary and cardiovascular tissue, BMC Syst Biol. 5: 168. 5 Westra, J.W. et al. (2013) Construction of a computable network model for non-diseased pulmonary tissue. Bioinformatics and biology insights 7:1-26. 6 Gebel, S. et al. (2013) Construction of a computable network model for DNA damage, autophagy. cell death, and sensecence. Bioinformatics and Biology Insights 7:97-117. 7 Park, J.S. et al. (2013) Construction of a computable network model of tissue repair and angigenesis in the lung. J. Clin. Toxicol., SI2, 002. 8 Martin, F. et al. (2014) Quantification of biological network perturbations for mechanistic insight and diagnostics using two-layer causal models. BMC Bioinformatics 15, 238. 9 Gaworski C.L. et al. (1987) 13-week inhalation toxicity study of menthol cigarette smoke. Food and chemical toxicology: 35, E83-682. 10 Vanscheeuwijck P. et al. (2012) Evaluation of the potential effects of ingredients added to cigarettes. Part 4: Food and chemical toxicology: 40(1):113-31. 11 Coggins C.R. et al. (2014) Toxicological assessment of kretek cigarettes Part 3: kretek and American-blended cigarettes. inhalation toxicity. Regulatory toxicology and pharmacology : RTP 70 Suppl 1, S26-40. 12 Piade J.J. et al. (2014) Toxicological assessment of kretek cigarettes Part 3: kretek and American-blended cigarettes. inhalation toxicity. Regulatory toxicology and pharmacology : RTP 70 Suppl 1, S26

PMI SCIENCE PHILIP MORRIS INTERNATIONAL



Competing Financial Interest

The research described in this poster was sponsored by Philip Morris International.