



PMI RESEARCH & DEVELOPMENT

IIVS Workshop on “Assessment of In Vitro COPD Models For  
Tobacco Regulatory Science”

*“ Combining Systems Biology, a Computational Approach  
and a Human Organotypic In Vitro Model Exposed to Whole  
Cigarette Smoke: An Example of 21<sup>st</sup> Century Toxicology  
Assessment”*

Carole Mathis, Philip Morris International, R&D

December 9<sup>th</sup>, 2014

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- Step 1: Characterization of Both *In Vitro* Model/Exposure System
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# Background

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2007: Publication by the U.S. National Research Council of a new strategy plan for toxicology assessment to update and advance our knowledge on the toxicity and the Mode of Action of environmental agents.

Example of recommended approaches:

- Medium and high-throughput *in vitro* screening assays
- Computational toxicology
- Systems biology
- Pharmacokinetic modeling

21st Century Toxicology

2009: European Commission published a report on «Alternative Testing Strategies for «Replacing, Reducing, and Refining» («3R») the use of animals in research.

- Human organotypic tissues based on primary cells cultured in three dimensions, with proper cell-cell contact, recapitulating biological functions (e.g. mucus secretion, muco-ciliary clearance,..) and allowing whole cigarette smoke exposure at the air-liquid interface.

# Background to Case Study

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- Smoking causes serious diseases such as cardiovascular diseases, lung cancer and chronic obstructive pulmonary disease.
- Philip Morris International is therefore developing novel products that may have the potential to reduce smoking-related disease risk compared to conventional combustible cigarettes.
- To determine whether such potentially reduced-risk products (RRP) have the potential to reduce disease risk, among the other things, we compare their biological impact with that of a combustible reference cigarette (3R4F) on a mechanism-by-mechanism basis.

# Quantitative Mechanism-Based Systems Impact Assessment

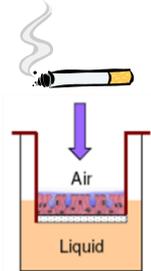
Experimental data production

Compute systems response profiles

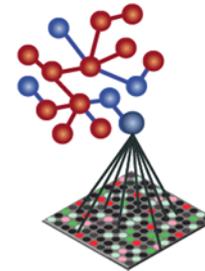
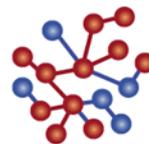
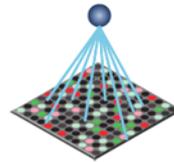
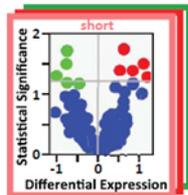
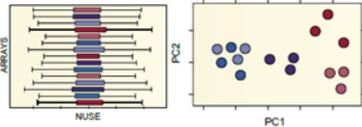
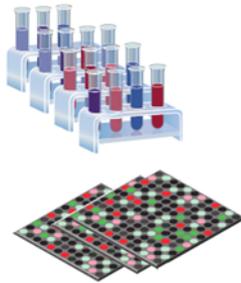
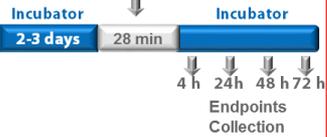
Identify perturbed biological networks

Compute network perturbation amplitudes

Compute product biological impact

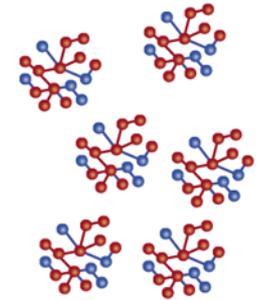
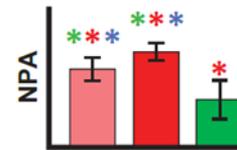


Whole smoke exposure

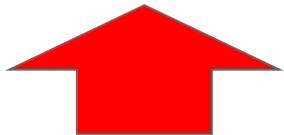
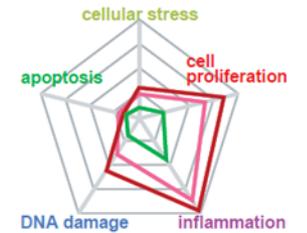


Find  $x$  that minimizes  $\sum_{\text{edges } (i,j)} \omega(i,j) (x_i - \text{sign}(i,j)x_j)^2$  subject to  $|x|_{\text{transcript layer}} = \beta$  where  $\omega(i,j)$  gives the edge weights.

Evaluate  $\sum_{\text{edges } (i,j)} (x_i + \text{sign}(i,j)x_j)^2$  where the sum includes all the edges of the functional layer.



For each SRP<sub>c</sub>, evaluate  $BIF(c) = \sum_{\text{networks } n} \omega(n,c) \cdot NPA(n,c)$  where the weights  $\omega(n,c)$  are non-zero for perturbed networks (\*\*\*) only.



# Step 1: Characterization of Both *In Vitro* Model/Exposure System

## Exposure system characterization:

- **Characterization of the Vitrocell® 24/48 in vitro aerosol exposure system using mainstream cigarette smoke**

*S Majeed, S Frentzel, S Wagner, D Kuehn, P Leroy, PA Guy, A Knorr, J Hoeng, MC Peitsch*  
Chemistry Central J. in press.

## Organotypic tissue cultures: How close are they to *in vivo*?

- **Human bronchial epithelial cells exposed in vitro to cigarette smoke at the air-liquid interface resemble bronchial epithelium from human smokers.**

*Mathis C, Poussin C, Weisensee D, Gebel S, Hengstermann A, Sewer A, Belcastro V, Xiang Y, Ansari S, Wagner S, Hoeng J, Peitsch MC.*

*Am J Physiol Lung Cell Mol Physiol.* 2013 Apr 1;304(7):L489-503.

- **Systems approaches evaluating the perturbation of xenobiotic metabolism in response to cigarette smoke exposure in nasal and bronchial tissues.**

*Iskandar AR, Martin F, Talikka M, Schlage WK, Kostadinova R, Mathis C, Hoeng J, Peitsch MC.*

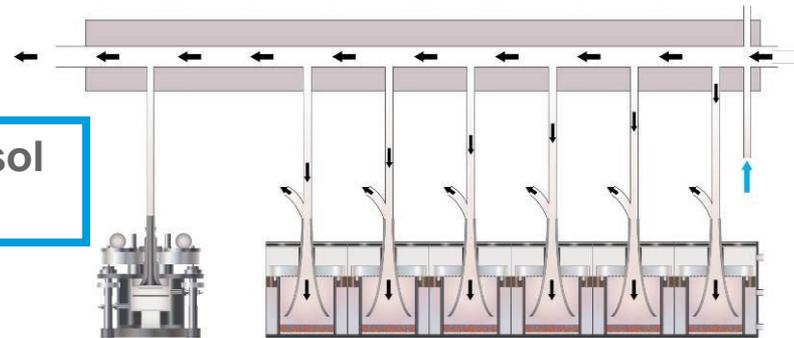
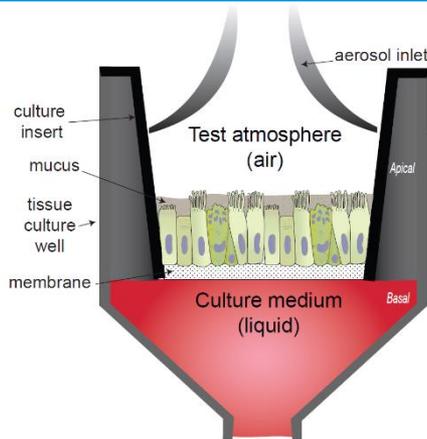
*Biomed Res Int.* 2013;2013:512086.

# VITROCELL® Whole Smoke Exposure System

## Climatic chamber with an exposure module

Up to 48 wells can be exposed simultaneously. The base module has a format of 8 rows x 6 columns.

The smoking machine produces aerosol from research cigarettes and RRP



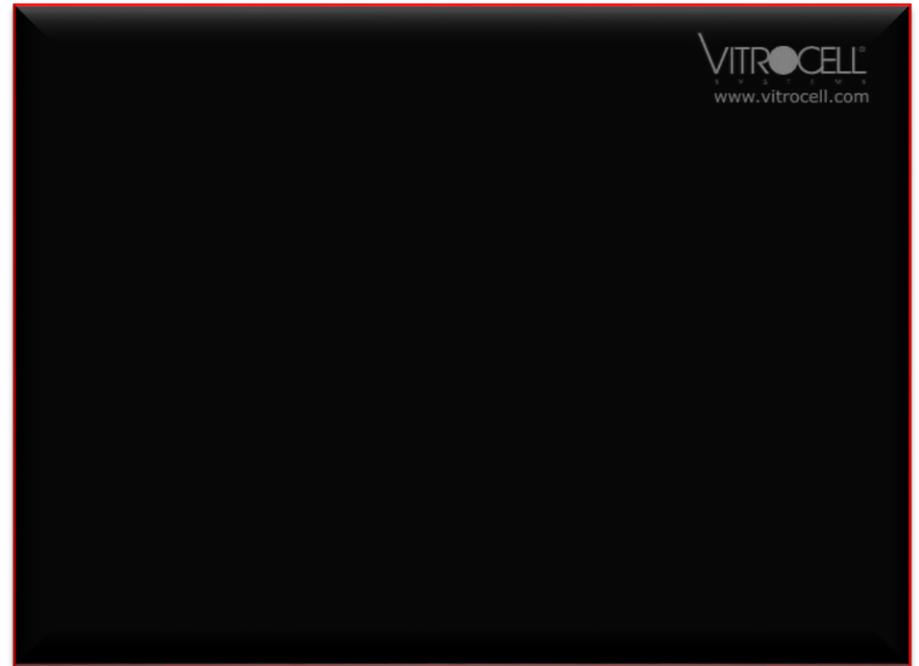
Human organotypic bronchial tissue cultures can be exposed at the air-liquid interface to whole mainstream smoke of the 3R4F or aerosol generated from RRP

# Whole Cigarette Smoke/Aerosol Exposure System (Vitrocell®)

VITROCELL® EXPOSURE SYSTEM

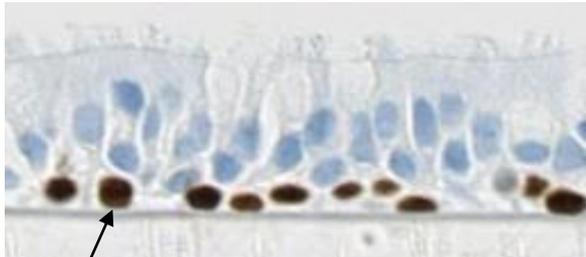


VITROCELL® DEPOSITION SENSOR



# Human Organotypic Bronchial Epithelial Cells Resemble *In Vivo* Bronchial Epithelium

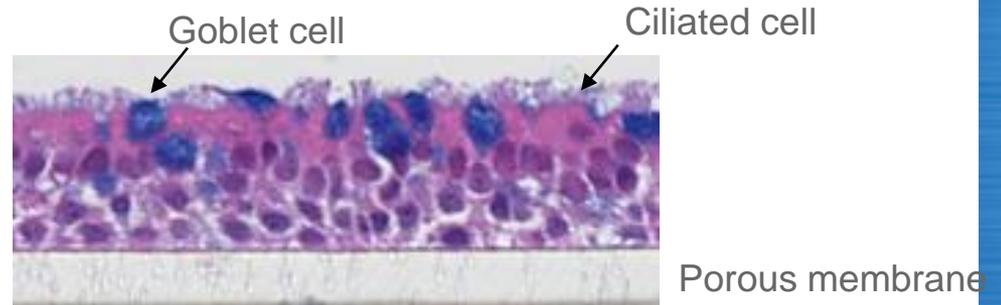
Apical



Basal cell

P63 immunostaining

Basal



Porous membrane

Hematoxylin/Eosin/Alcian Blue

Unperturbed human organotypic bronchial epithelial cell culture closely resembles to human lung epithelium both at the morphological level (Karp et al. 2002) and at the molecular level (Pezzulo et al. 2011).



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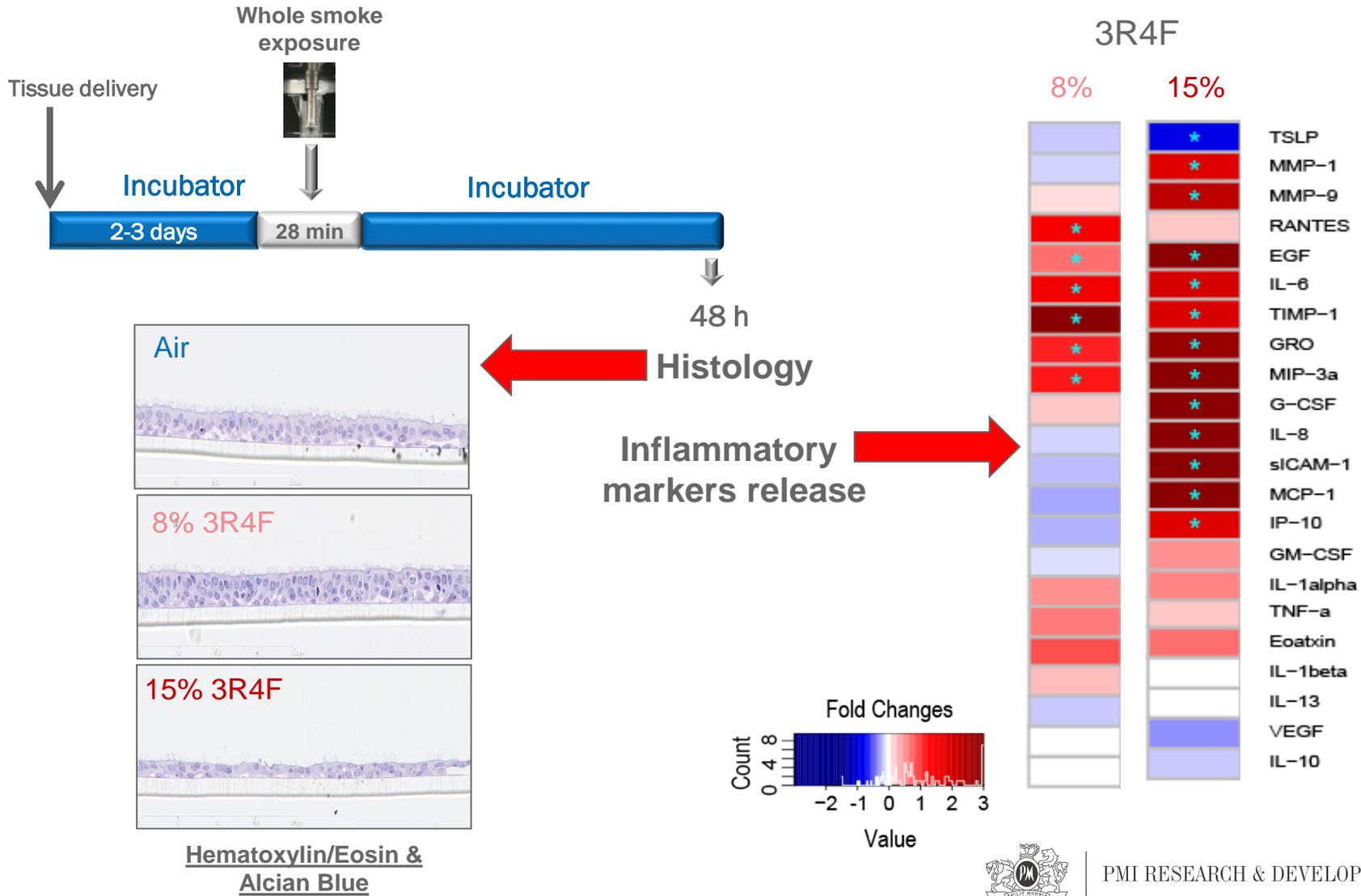
# Goblet Cells Metaplasia Can Be Induced in Human Bronchial Organotypic Tissue Cultures

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- **HBEC cultured in ALI system in the presence of IL-13 (10ng/mL, from day 2) and a combination of IL-13 and IL-4 (10ng/mL, days 6-12) developed mucous cell metaplasia.** – *Eicosanoid biosynthesis during mucociliary and mucous metaplastic differentiation of bronchial epithelial cells* - Jakiela B. et al. Prostaglandins and other Lipid Mediators; 106 (2013) 116-123.
- **Mucous metaplasia is induced in primary human airway epithelial cells cultured at ALI and treated with IL-13 for 5 days.** – *SAM-pointed domain ETS factor mediates epithelial cell-intrinsic innate immune signaling during airway mucous metaplasia* – Korfhagen, TR. et al. Proc. Natl. Acad. Sci. USA (2012) Oct 9;109(41):16630-5.
- **IL-4 induces mucous cell metaplasia in primary human bronchial epithelial cells cultured at ALI.** – *Association of TMEM16A chloride channel overexpression with airway goblet cell metaplasia* – Scudieri P. et al. J. Physiol. (2012) 590-23: 6141-6155.



# Example of CS Acute Exposure Impact on Human Organotypic Bronchial Epithelial Cells



# Organotypic Cultures of Human Primary Bronchial Epithelial Cells Exposed to Whole Smoke

Experimental  
Data  
Production

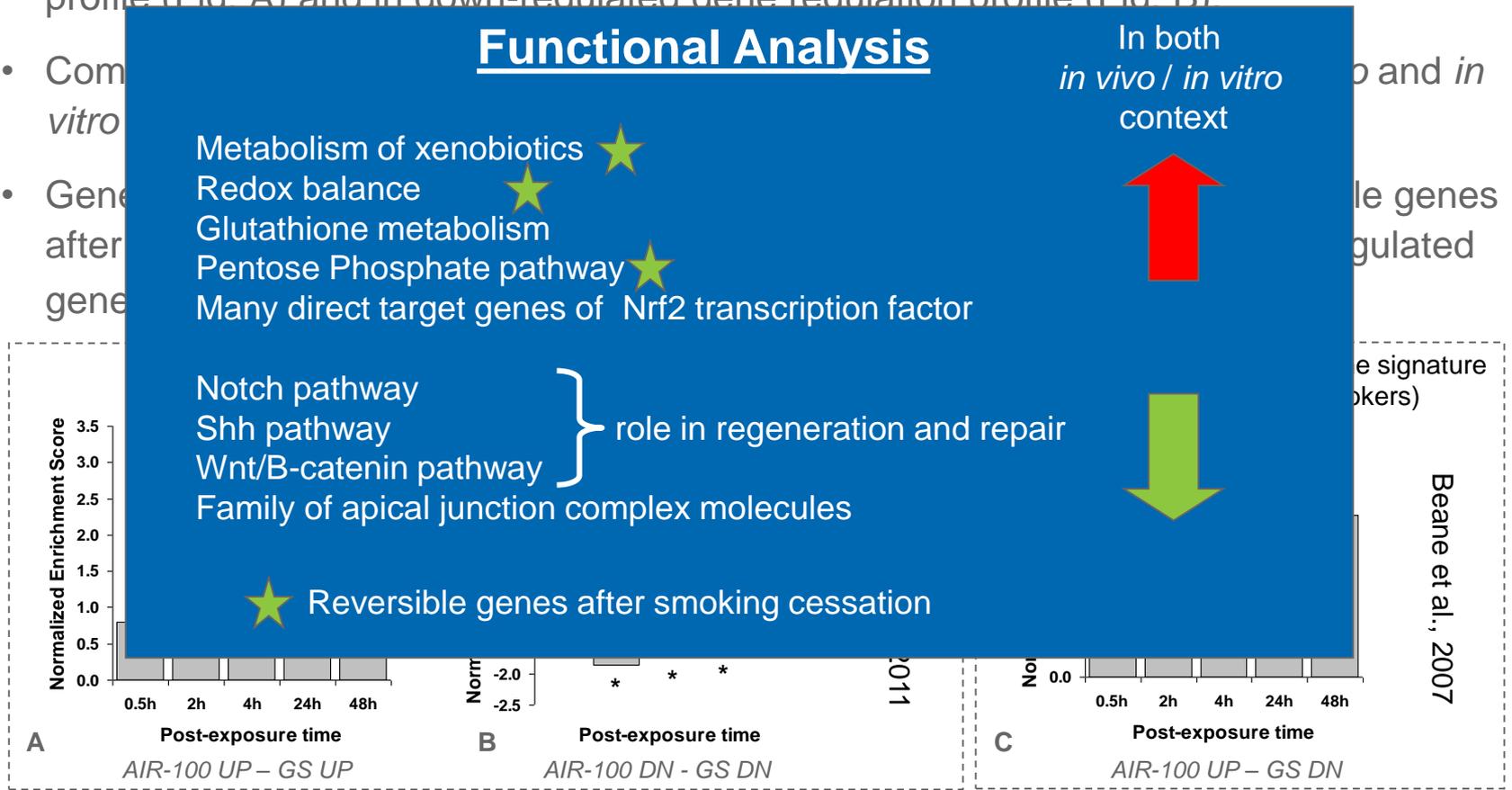
		<u>Experimental Design</u>								
TEST SUBSTANCE		SHAM				CIGARETTE SMOKE				ENDPOINTS
Exposure Time (Min)	7	14	21	28	7	14	21	28	Gene Expression MicroRNA MMP-1 Release Differential Cell Counts Survival	
	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5		
Post-Exposure (p-e) Time (Hrs)	2	2	2	2	2	2	2	2		
	4	4	4	4	4	4	4	4		
	24	24	24	24	24	24	24	24		
	48	48	48	48	48	48	48	48		



# Human Organotypic Bronchial Epithelial Cells Exposed to CS Resemble Bronchial Epithelium From Human Smokers

- For all four *in vivo* smoking gene signatures used in the GSEA, a similar pattern of enrichment score was found in CS-exposed AIR-100 up-regulated gene regulation profile (Fig. A) and in down-regulated gene regulation profile (Fig. B)

- Common to both *in vitro* and *in vivo* contexts
- Gene expression profiles after 48h of CS exposure resemble gene expression profiles of human smokers

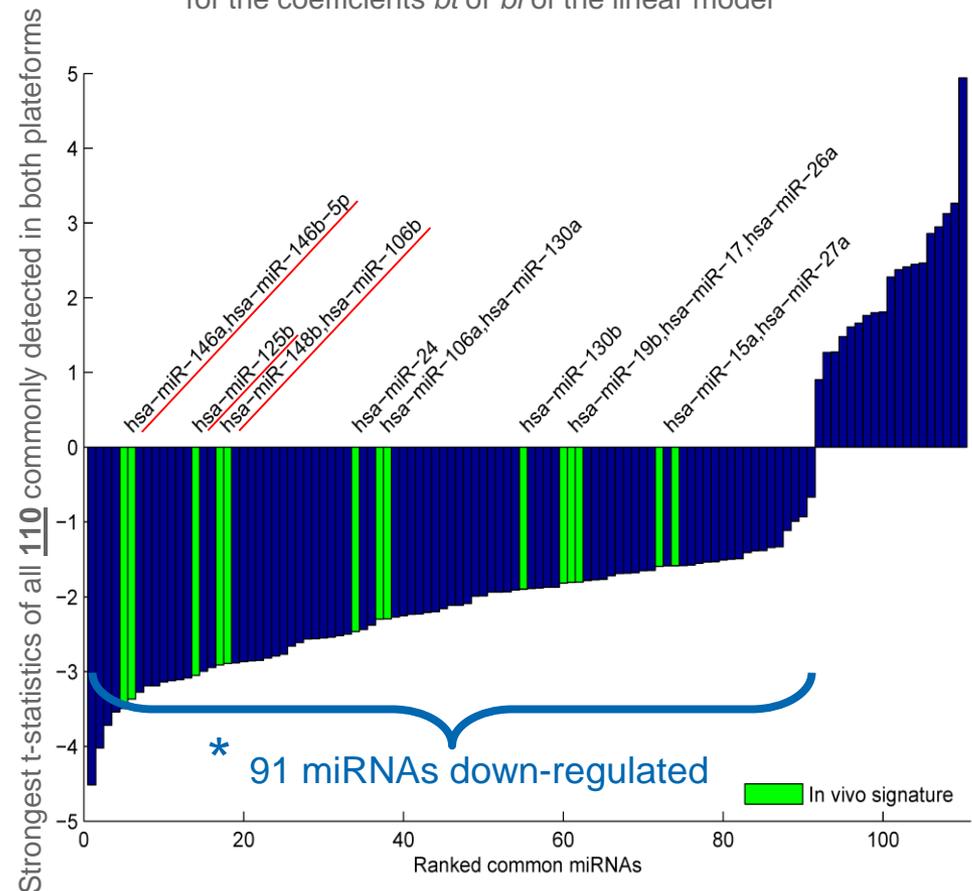


# Human Organotypic Bronchial Epithelial Cells Exposed to CS Resemble Bronchial Epithelium From Human Smokers

- Only one human *in vivo* miRNA dataset from bronchial epithelial cells published so far (Schembri et al. 2009).
- Out of ~ 230 miRNAs detectable in this tissue context, half of them are commonly detected in both *in vivo* and our *in vitro* studies. Only 14 miRNAs differentially expressed are common between both *in vivo* and *in vitro* datasets (GREEN tag).
- CS down-regulates a large majority of miRNA expression (\*: 91 miRNAs out of 110) in both *in vivo* and *in vitro* situation.
- The biological functions associated with some of the highly “translatable” miRNAs are related to inflammation (miR-146b and miR-125b) and cell cycle processes (miR-106a and miR-106b) that are also known to be perturbed by CS in lung tissue context.

## Comparison between *in vivo* human smoker miRNA signature and CS-exposed AIR-100 *in vitro* miRNA dataset.

The vertical axis of the bar plot represents the best t-statistics (i.e. lowest negative or highest positive t-scores) obtained for the coefficients *bt* or *bi* of the linear model



# Advantages/Limitations of the Air-Lifted Bronchial Epithelial Culture Model

## Advantages

- Human primary cells (No species translatability issue - 3 Rs)
- Different donors available (e.g., smokers, non smokers, COPD patients)
- Long-term culture possible
- Like *in vivo*:
  - Direct exposure to whole smoke at the air-liquid interface
  - Similar morphology/structure (Goblet cells, basal cells, ciliated cells, tight junctions, pseudostratified epithelium)
  - Similar gene expression pattern (normal untreated conditions)
- Various endpoints can be collected:
  - From tissue insert: RNA/miRNA/DNA/Proteins/Metabolites, morphological changes (histo/IHC), Tissue integrity (TEER), ion channels activity measurement
  - From basal side medium: Release of inflammatory markers, cytotoxicity (AK assay, LDH release,..), CYPs activity
  - From the apical side: Mucin secretion, cilia beating frequency, Mucociliary clearance

## Limitations

- Limited number of inserts from one donor
- Risk of contamination when performing chronic/long-term exposure
- Donor-to-donor variability
- Missing other key players (immune cells, fibroblasts, smooth muscle cells)
- Difficult to find the right exposure design (appropriate dose, how many exposure per day/per week, reproducibility...) -> Large experiment -> High cost/Time consuming

# Step 2: Mono- or Co-culture Organotypic Bronchial Tissue Culture?

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## **An Impact Assessment of Cigarette Smoke on Organotypic Models of Bronchial Epithelial Mono-culture and Bronchial Epithelial/Fibroblast Co-culture**

*A Iskandar, X Yang, S Frenzel, C Mathis, P Leroy, D Kuehn, S Majeed, C Merg, A Elamin, E Guedj, R Dulize, F Martin, M Talikka, MC Peitsch and J Hoeng*

POSTER

# Step 3: Assessment of Whole Smoke Exposure Impact on Organotypic Bronchial Tissue Culture

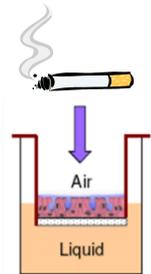
Experimental data production

Compute systems response profiles

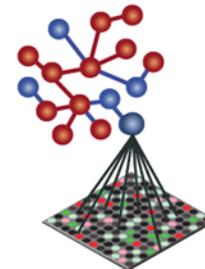
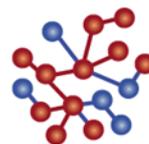
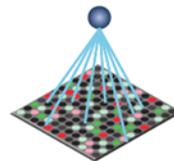
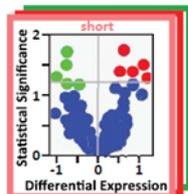
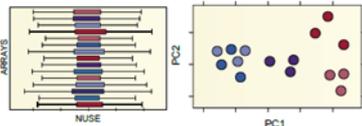
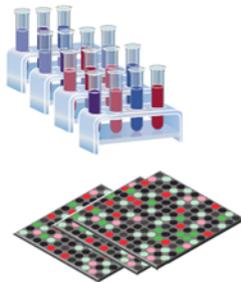
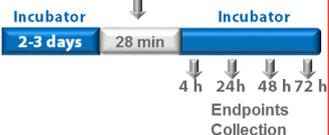
Identify perturbed biological networks

Compute network perturbation amplitudes

Compute product biological impact

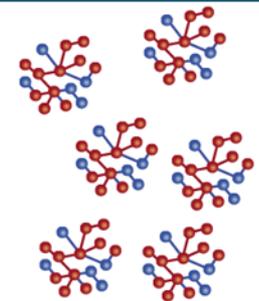
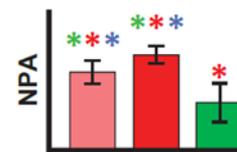


Whole smoke exposure

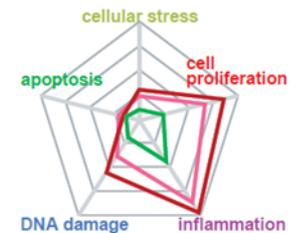


Find  $x$  that minimizes  $\sum_{\text{edges } (i,j)} \omega(i,j) (x_i - \text{sign}(i,j)x_j)^2$  subject to  $|x|_{\text{transcript layer}} = \beta$  where  $\omega(i,j)$  gives the edge weights.

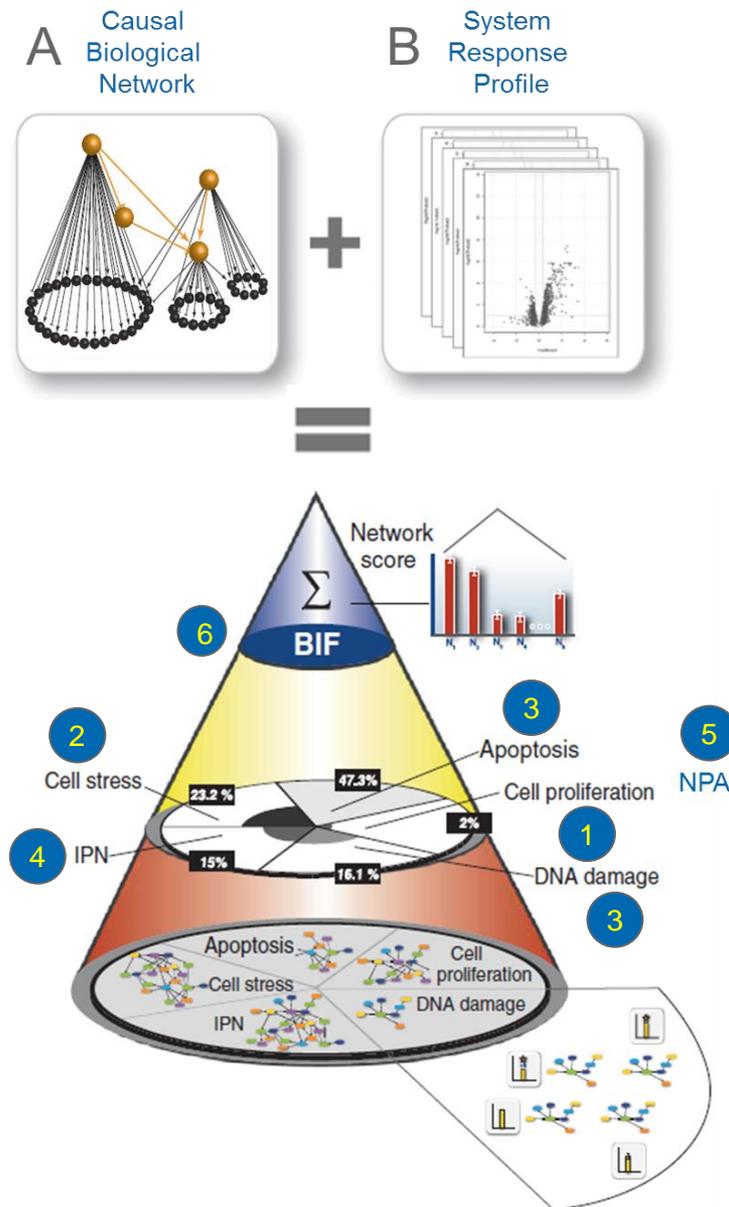
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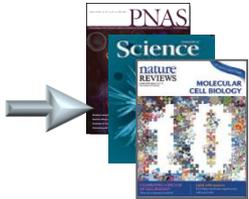
# Systems Toxicology Assessment of Whole Smoke Exposure



- 1. Construction of a computable cell proliferation network for non-diseased lung tissue.** *J. Westra, et al. BMC Systems Biology, 2011 Jul 5:105.*
- 2. Construction of a computable cellular stress network for non-diseased lung and cardiovascular tissue.** *W.K. Schlage, et al. BMC Systems Biology, 2011 Oct 5:168.*
- 3. Construction of a Computable Network Model for DNA Damage, Cell Death, Autophagy, and Senescence.** *S. Gebel, et al. Bioinformatics and Biology Insights 2013 7:97-117.*
- 4. A modular cell-type focused inflammatory process network model for non-diseased pulmonary tissue.** *J. Westra, et al. Bioinformatics and Biology Insights 2013 Jun 20; 7:167-92.*
- 5. Assessment of network perturbation amplitude by applying high-throughput data to causal biological networks.** *F. Martin, et al. BMC Systems Biology 2012, 6:54.*
- 6. Quantification of biological network perturbations for mechanistic insight and diagnostics using two-layer causal models assessment of biological impact using transcriptomic data and mechanistic network models.** *F. Martin, et al. BMC Bioinformatics. 2014 Jul 11;15(1):238.*



# Biological Statements from Scientific Literature/Original Research Are Coded into Network Models Using Biological Expression Language (BEL)

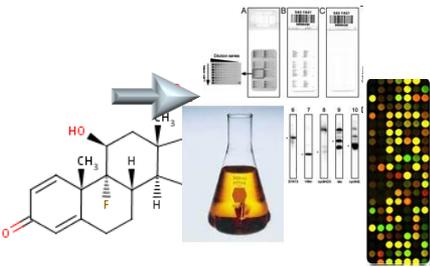


Scientific Literature

“RNA expression of RBL2 is directly mediated via activation of the FOXO3 transcription factor”

`tscript(p(HGNC:FOXO3)) => r(HGNC:RBL2)`

*J Biol Chem* 2002 Nov 22 277(47) 45276-84



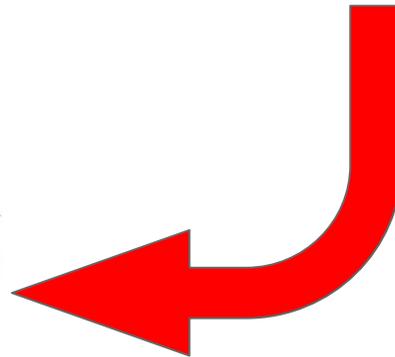
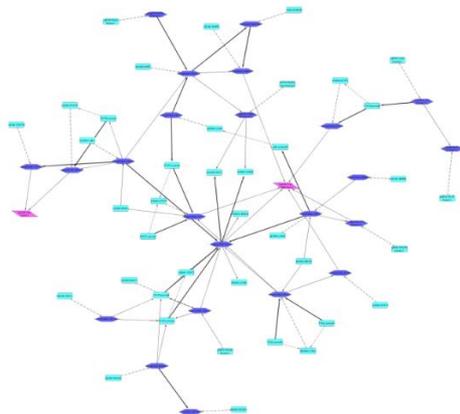
Original Research

“LY294002 inhibits the activity of the PI3K alpha catalytic subunit”

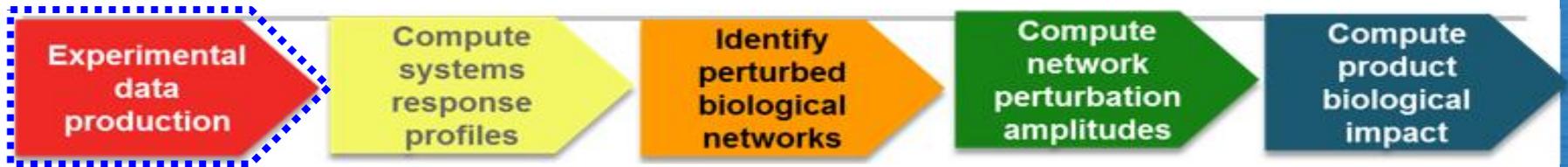
`a(CHEBI:LY294002) -| kin(p(HGNC:PIK3CA))`

XYZ Corp Document 12345

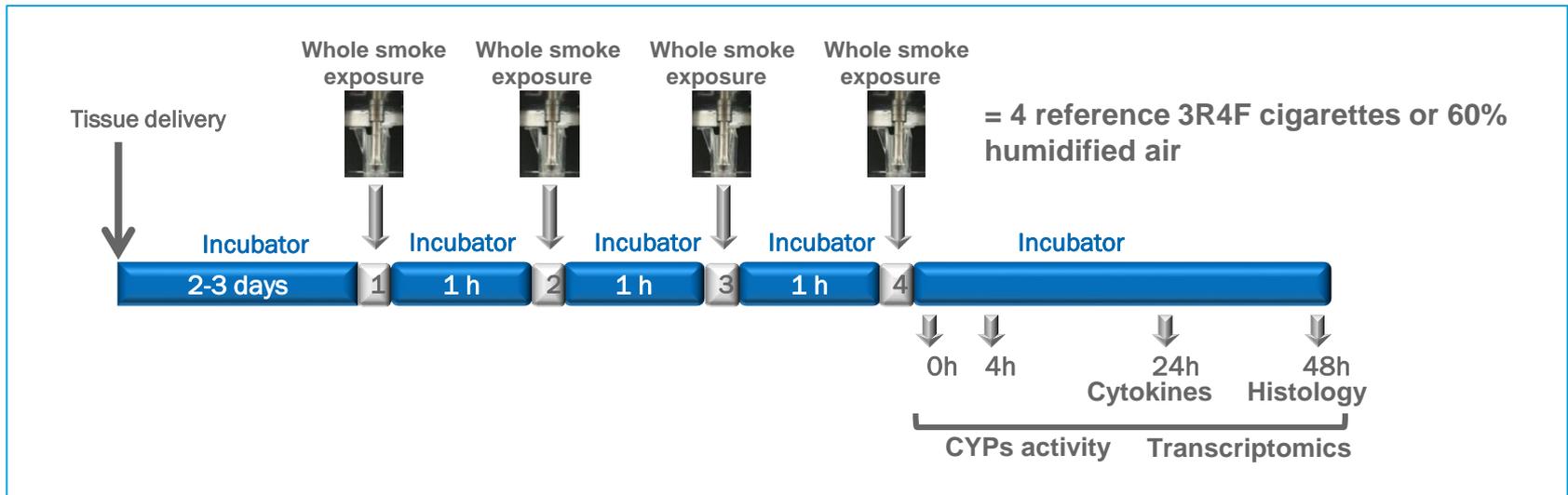
Network Models



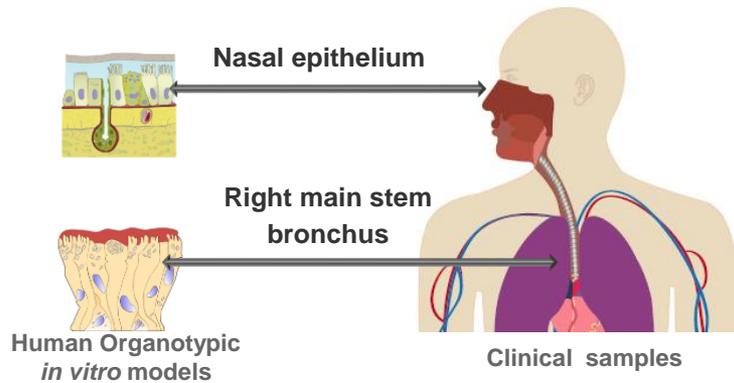
# Repeated Whole Smoke Exposure of Organotypic Cultures Derived from Human Primary Epithelial Cells



## EXPERIMENTAL DESIGN



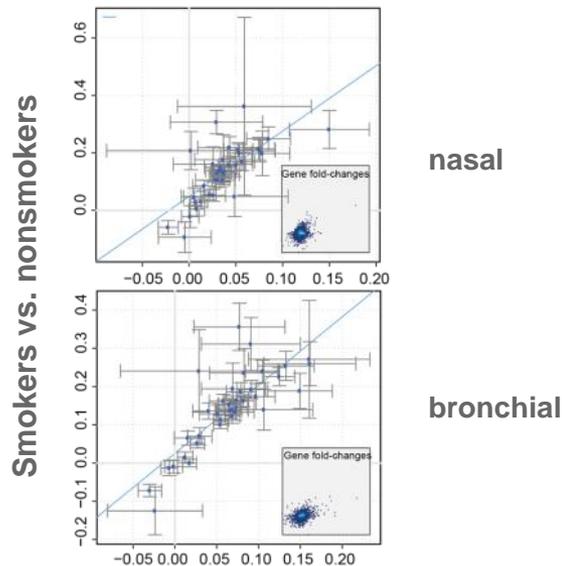
# Comparison of Clinical Samples from Smokers to CS Exposed Nasal and Bronchial Organotypic Cultures



Healthy Non-smoker (n=14) Age	Healthy Smoker (n=13) Age	Smoker Pack-years
31.6 ± 10.8	35.4 ± 9.9	10.77 ± 9.3

(Zhang X. et al., *Physiol. Genomics* 2010)

## Perturbation of the Xenobiotic metabolism network model

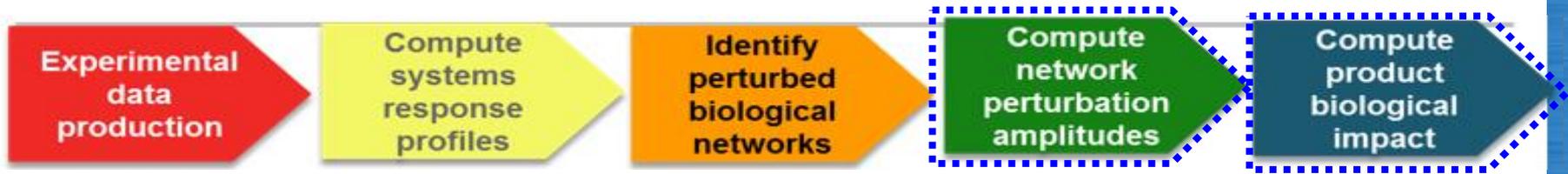


CS vs. air exposed organotypic

**Systems Approaches Evaluating the Perturbation of Xenobiotic Metabolism in Response to Cigarette Smoke Exposure in Nasal and Bronchial Tissues**

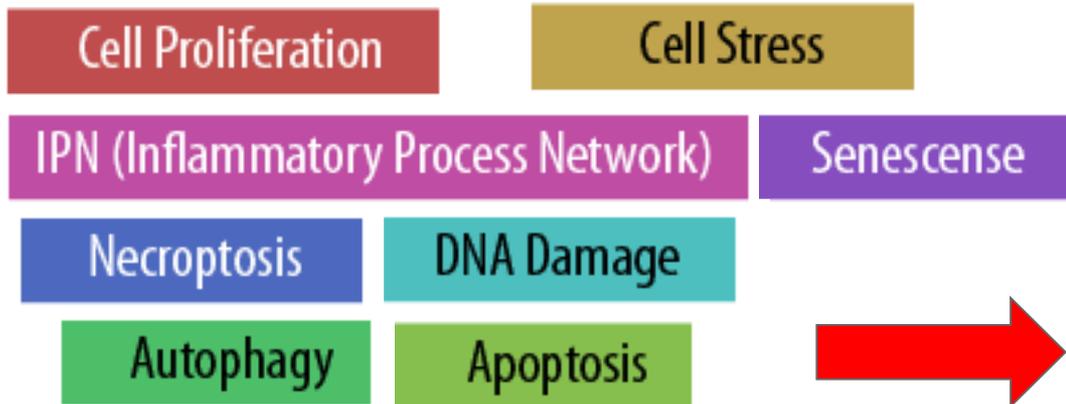
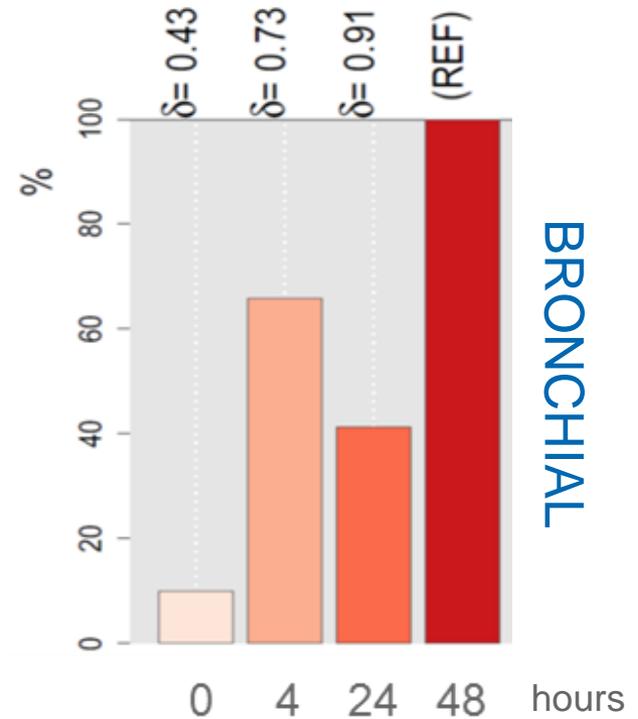
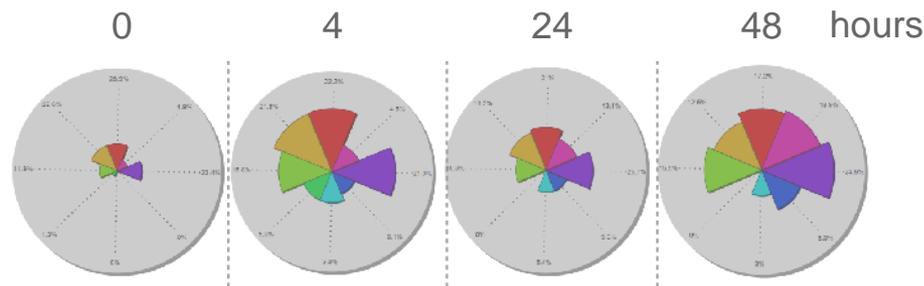
*Iskandar AR, Martin F, Talikka M, Schlage WK, Kostadinova R, Mathis C, Hoeng J, Peitsch MC. Biomed Res Int. 2013 Oct 3*

# Example of 3R4F Repeated Exposure Impact Quantification



## Network Perturbation Amplitude

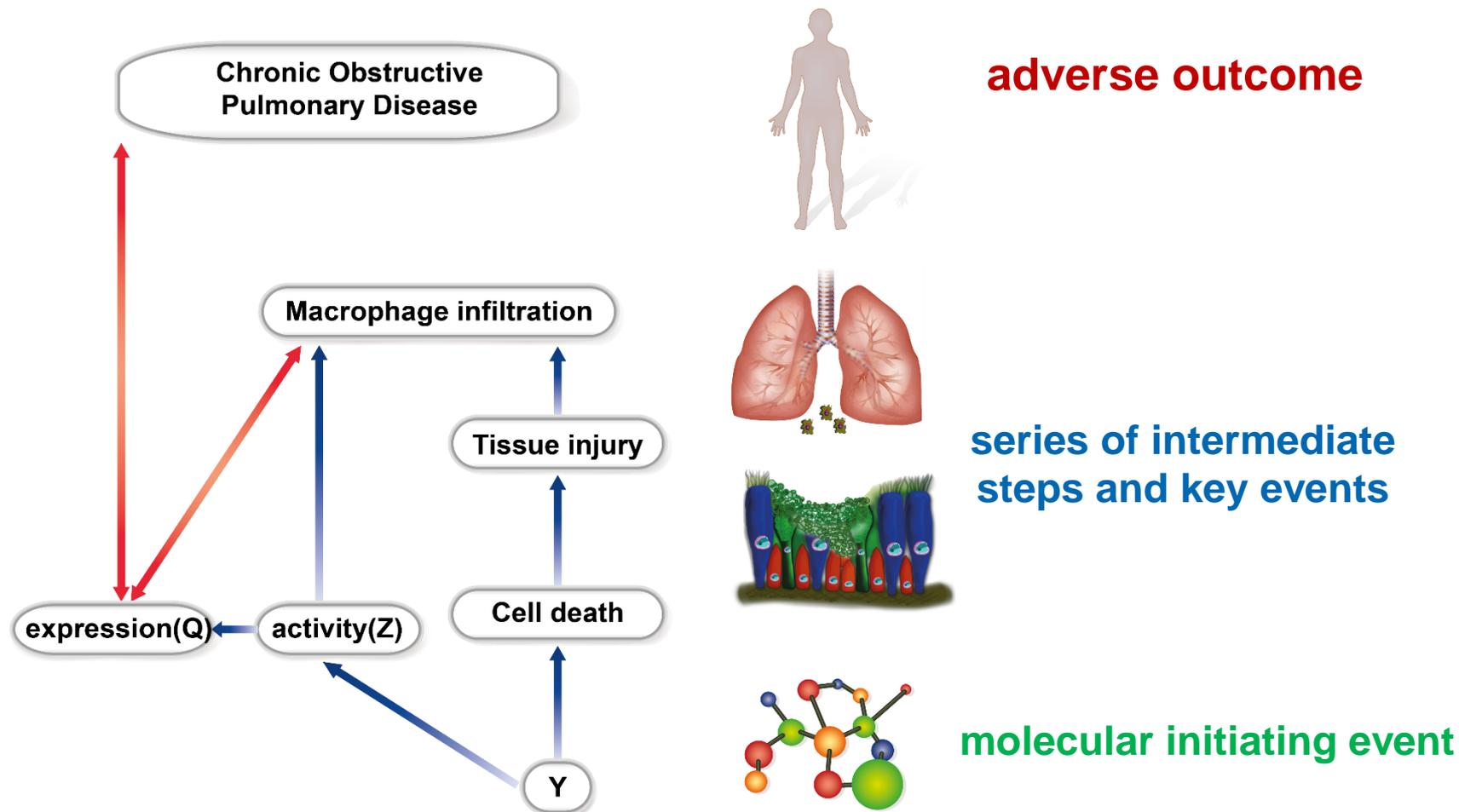
## Relative BIF



17% 3R4F (Nicotine 0.24 mg/L)

# Network Models to Study Adverse Outcome Pathways

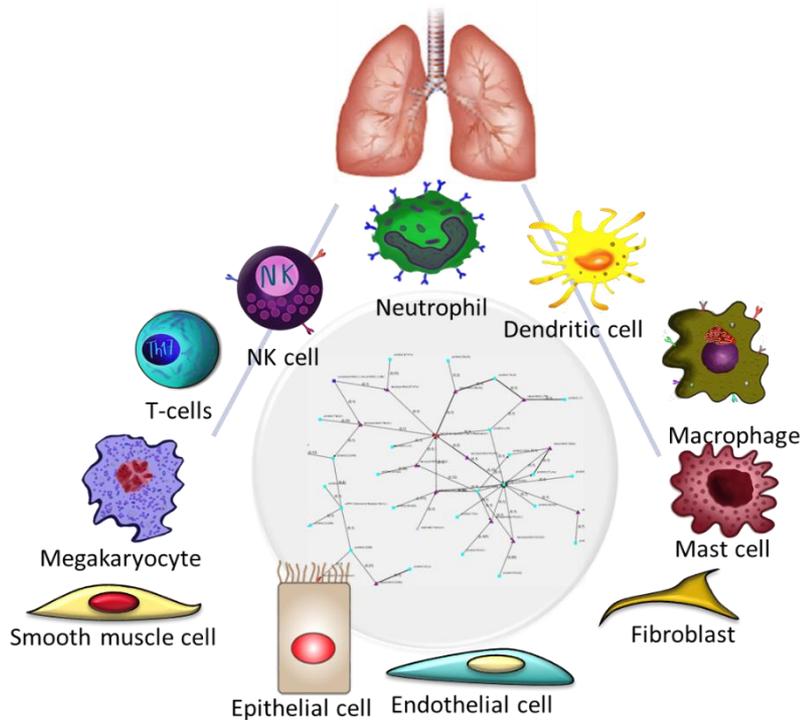
Network model gives “a mechanistic representation of critical toxicological effects that span over different layers of biological organization”



Vinken M (2013) The adverse outcome pathway concept: a pragmatic tool in toxicology. *Toxicology* 312:158-165  
Vinken M, Whelan M, Rogiers V (2014) Adverse outcome pathways: hype or hope? *Archives of toxicology* 88:1-2

# NETWORK VERIFICATION CHALLENGE

The purpose of the Network Verification Challenge is to engage the scientific community to review, challenge, and make corrections to the conventional wisdom.



- Mucus hypersecretion**  
**Epithelial cell proinflammatory signaling**  
**Epithelial barrier defense**
- Microvascular endothelium activation**
- Macrophage activation**  
**Macrophage differentiation**  
**Macrophage mediated neutrophil recruitment**
- Neutrophil response**  
**Neutrophil chemotaxis**
- Dendritic cell activation**  
**Dendritic cell migration to tissue**  
**Dendritic cell migration to lymph**
- Th1 differentiation**      **Th1 response**  
**Th2 differentiation**      **Th2 response**  
**Th17 differentiation**      **Th17 response**  
**Tc response**                  **Treg response**
- Mast cell activation**
- NK cell activation**
- Megakaryocyte differentiation**
- Tissue damage**



The sbv IMPROVER project, the website and the Symposia are part of a collaborative project designed to enable scientists to learn about and contribute to the development of a new crowd sourcing method for verification of scientific data and results. The current challenges, website and biological network models were developed and are maintained as part of a collaboration with Selventa, OrangeBus and ADS. The project is funded by Philip Morris International.



# Network Verification Challenge in a Nutshell

## 1. Network Model Construction

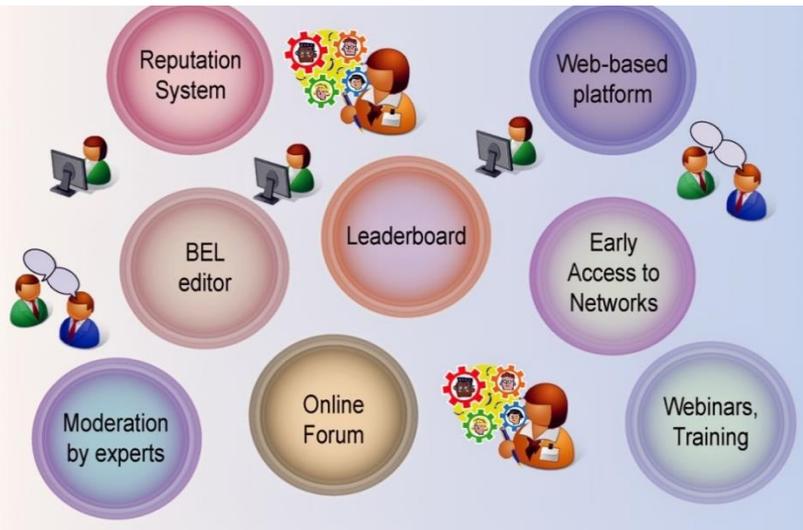
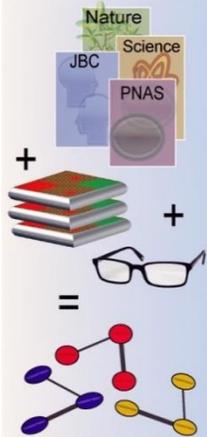
## 2. Online Crowd-Verification

## 3. Jamboree Preparation

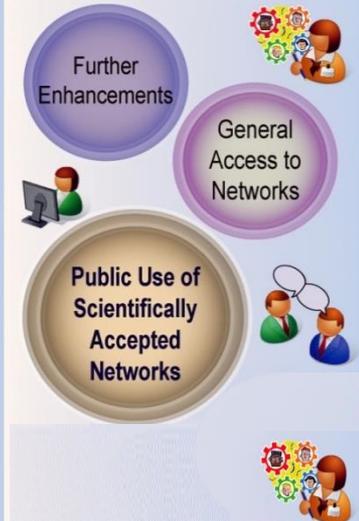
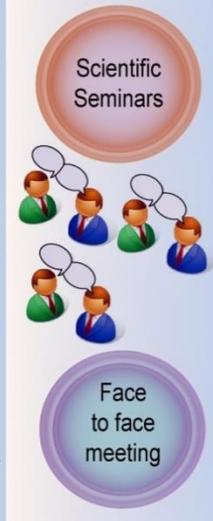
## 4. Network Jamboree

## 5. Network Dissemination Continuous Improvement

Boundary Definition +



Scoring  
Analysis  
Prioritization  
Evidences Summary



The sbv IMPROVER project team (2013). On Crowd-verification of Biological Networks. **Bioinformatics and Biology Insights** 2013:7 307-325.

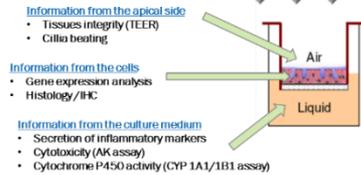


PMI RESEARCH & DEVELOPMENT

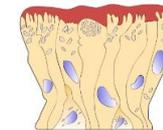
# Acknowledgments



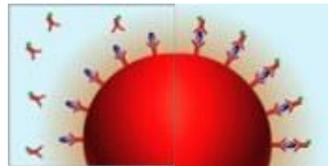
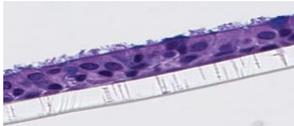
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