

Filippo Zanetti^a, Bjoern Titz^a, Alain Sewer^a, Giuseppe Lo Sasso^a, Elena Scotti^a, Walter K. Schlage^b, Carole Mathis^a, Patrice Leroy^a, Shoab Majeed^a, Laura Ortega Torres^a, Brian R. Keppler^c, Ashraf Elamin^a, Keyur Trivedi^a, Emmanuel Guedj^a, Florian Martin^a, Stefan Frentzel^a, Nikolai V. Ivanov^a, Manuel C. Peitsch^a, Julia Hoeng^a

^a Philip Morris International Research and Development, Philip Morris Products S.A., Quai Jeanrenaud 5, 2000 Neuchâtel, Switzerland (part of Philip Morris International group of companies); ^b Biology consultant, Max-Baermann-Str. 21, 51429 Bergisch Gladbach, Germany; ^c Metabolon Inc., 617 Davis Drive, Suite 400, Durham, NC, USA

SUMMARY

Introduction. Smoking is one of the major lifestyle-related risk factors for periodontal diseases. Smoking can affect the structure of the epithelial mucosa, impair the inflammatory response, and change the redox status of the oral cavity [1]. Harm reduction through the development of Modified Risk Tobacco Products (MRTP) provides a promising opportunity for adult smokers who would otherwise continue cigarette smoking. An MRTP is defined by the U.S. Family Smoking Prevention and Tobacco Control Act as “any tobacco product that is sold or distributed for use to reduce harm or the risk of tobacco related disease associated with commercially marketed tobacco products”. The Tobacco Heating System (THS) 2.2 is a candidate MRTP that uses a precisely controlled heating device into which a specially designed tobacco product, the Tobacco Stick, is inserted and heated to generate an aerosol [2].

Objectives. The objective was to assess – using a systems toxicology approach – how aerosol from a heat-not-burn technology-based candidate MRTP, THS2.2, compared to reference (3R4F) cigarette smoke (CS), affects human gingival epithelial organotypic cultures.

Materials & methods. Human gingival epithelial organotypic cultures (EpiGingiva™; MatTek) were repeatedly exposed (3 days) for 28 min to CS or THS2.2 aerosol at two concentration levels with similar nicotine content. Measured endpoints included histology, cytotoxicity, release of proinflammatory mediators, transcriptomics (mRNA and miRNA), and metabolomics. Effects on the transcriptome were assessed by gene-set and causal network analysis.

Results. Minor histopathological alterations and minimal cytotoxicity were observed upon THS2.2 aerosol exposure, while marked toxicity was observed for CS. Causal network and gene-set analysis of the transcriptomics data supported lower biological effects of THS2.2 aerosol than CS (e.g., with a ~79% reduction in the biological impact factor for the high concentrations). This included reduced effects of THS2.2 aerosol on oxidative stress, xenobiotic metabolism, and inflammation-related processes. Metabolomics confirmed that THS2.2 aerosol exposure was associated with lower oxidative stress than CS. In support of lower effects on inflammation-related processes, THS2.2 aerosol exposure resulted in a lower release of proinflammatory mediators than CS.

EXPERIMENTAL DESIGN / EXPOSURE

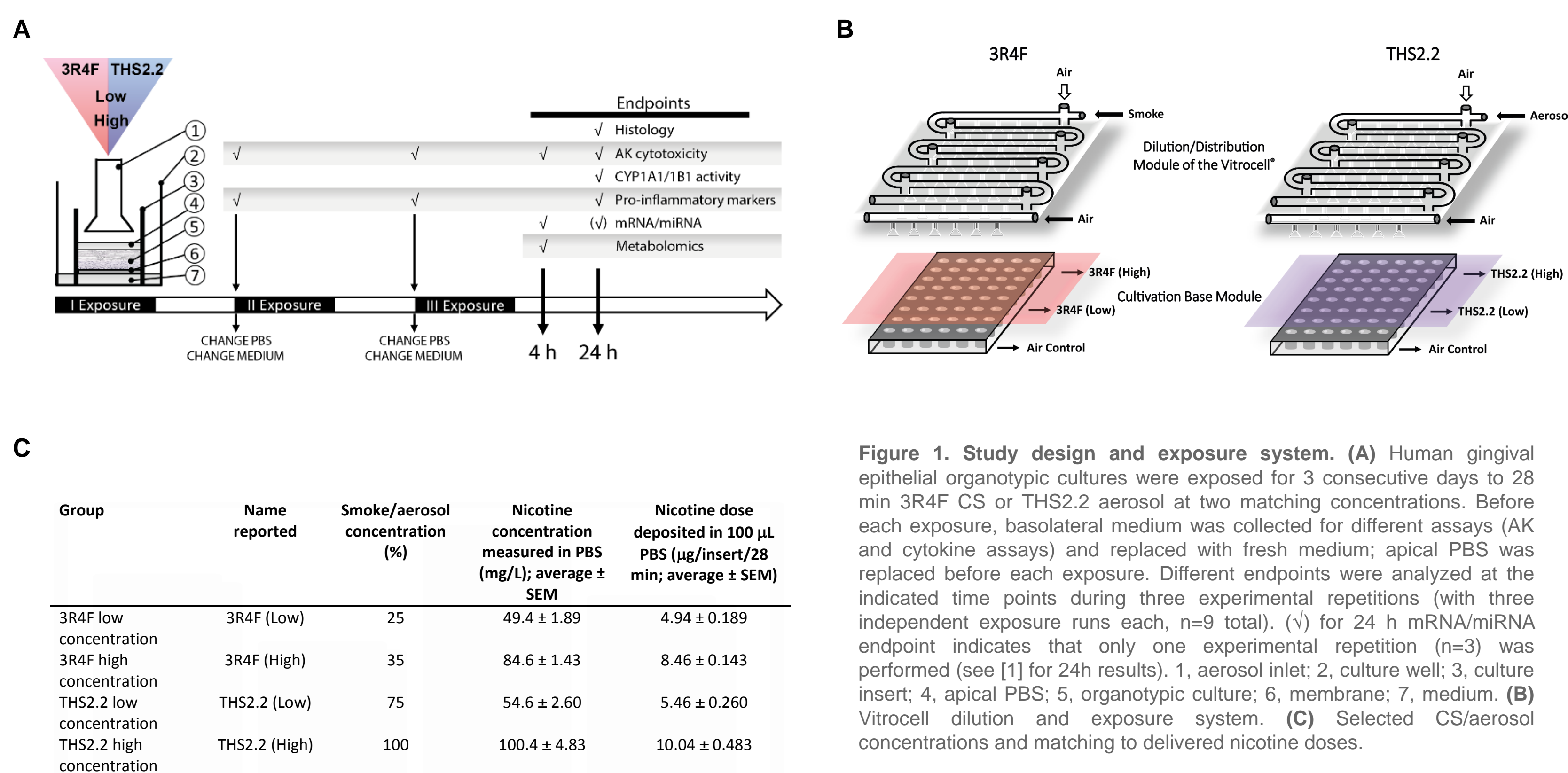


Figure 1. Study design and exposure system. (A) Human gingival epithelial organotypic cultures were exposed for 3 consecutive days to 28 min 3R4F CS or THS2.2 aerosol at two matching concentrations. Before each exposure, basolateral medium was collected for different assays (AK and cytokine assays) and replaced with fresh medium; apical PBS was replaced before each exposure. Different endpoints were analyzed at the indicated time points during three experimental repetitions (with three independent exposure runs each, n=9). (V) For 24 h mRNA/miRNA endpoint indicates that only one experimental repetition (n=3) was performed (see [1] for 24h results). 1, aerosol inlet; 2, culture well; 3, culture insert; 4, apical PBS; 5, organotypic culture; 6, membrane; 7, medium. (B) Vitrocult dilution and exposure system. (C) Selected CS/aerosol concentrations and matching to delivered nicotine doses.

RESULTS

CYTOTOXICITY AND TISSUE MORPHOLOGY

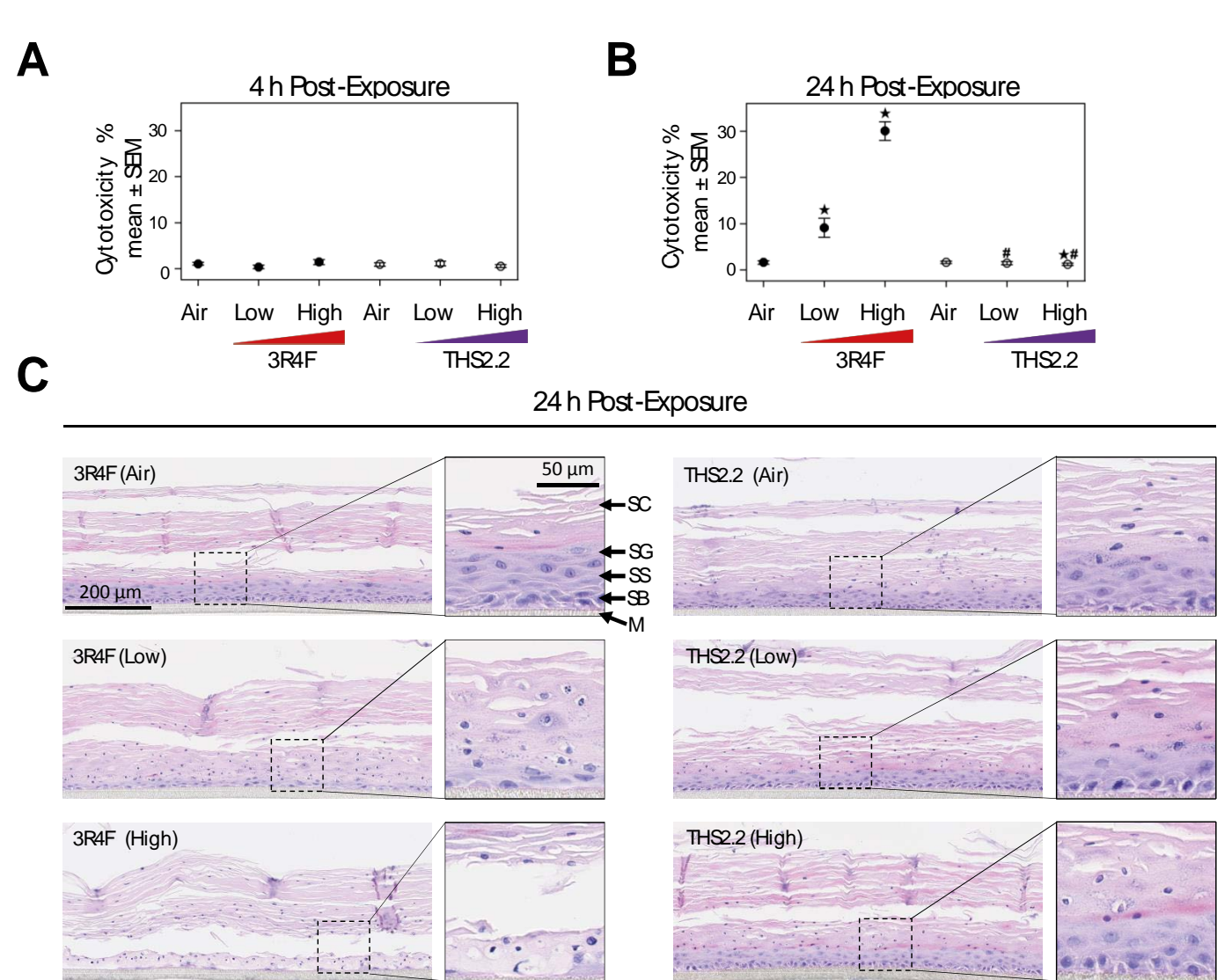


Figure 2. Cytotoxicity and tissue morphology of organotypic gingival cultures exposed to 3R4F CS and THS2.2 aerosol. Mean cumulative cytotoxicity levels were assessed using the AK assay at 4 h (A) and 24 h (B) post exposure time points. AK levels were normalized to values of the positive control (Triton-X-treated cultures, considered 100% cytotoxicity). Error bars indicate SEM (n=9). * p<0.05, compared with the corresponding air control; # p<0.05, compared with matching concentrations of 3R4F CS. (C) Representative images of H&E-stained gingival cultures after 24 h from the last exposure to 3R4F CS (left) or THS2.2 aerosol (right). Abbreviations indicate different layers of gingival cultures: M, membrane; SB, stratum basale; SS, stratum spinosum; SG, stratum granulosum; SC, stratum corneum. H&E images show 20x magnification, and 63x magnification for image insets. n=9.

OXIDATIVE STRESS

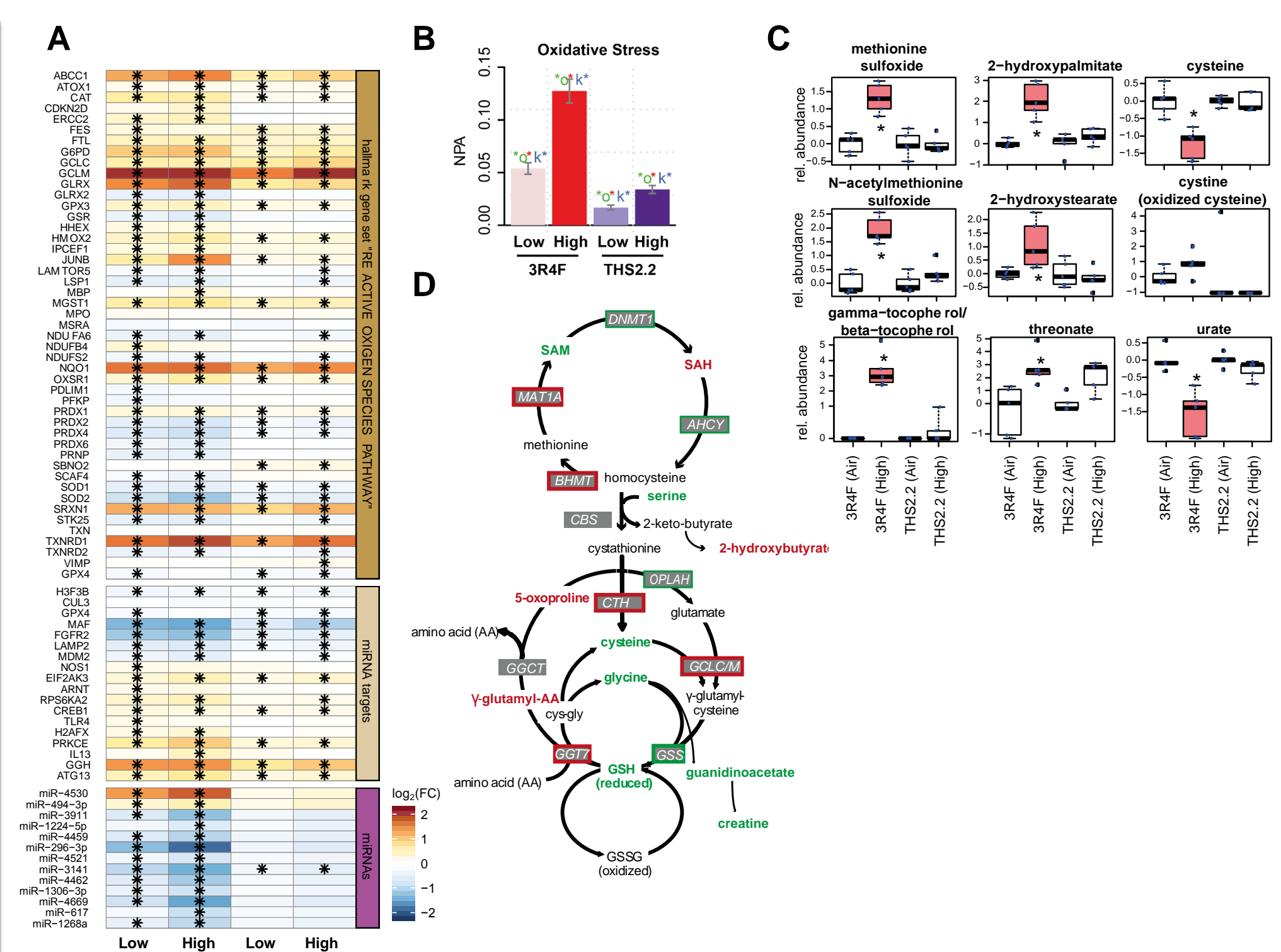


Figure 4. Differential induction of oxidative stress by 3R4F CS and THS2.2 aerosol. (A) Induction of oxidative stress response program: differential expression heatmap for genes of the reactive oxygen species pathway (HALLMARK_REACTIVE_OXIGEN_SPECIES_PATHWAY; M5938) (software.broadinstitute.org/gsea/msigdb), as well as for genes and miRNAs belonging to the “Oxidative Stress” candidate miRNA-mRNA network (see [1] for details). The “***” indicates statistically significant differential expression (FDR <0.05), n=6-9. (B) Assessment of exposure effects on the “Oxidative Stress” network. Bars show overall network perturbation amplitudes (NPA scores) based on transcriptomics data. Error bars delimit their 95% confidence intervals. Three statistics are shown: the red star indicates statistical significance with respect to the biological replication (i.e., 95% confidence intervals do not contain the 0 value), while the green and red stars indicate significant statistics with respect to the network structure (“O” and “K” statistics), n=6-9. (C) Metabolomics profiling was conducted 4 h after exposure of the tissue to high 3R4F CS and THS2.2 aerosol concentrations. Box plots summarize the response of metabolites sensitive to oxidative stress (blue dots indicate individual samples, n=5). Significant differences between exposed groups and their respective sham groups are indicated by filled colored boxes and a star (“***” means FDR <0.05). (D) Summary of exposure effects on glutathione and related metabolic reactions (see [1] for details). Relevant metabolic reactions of the glutathione pathway, including the gamma-glutamyl cycle, cysteine and methionine metabolism, and glycine, serine, and threonine metabolism. Significantly up- or down-regulated metabolites and genes are marked with red and green, respectively.

INFLAMMATION

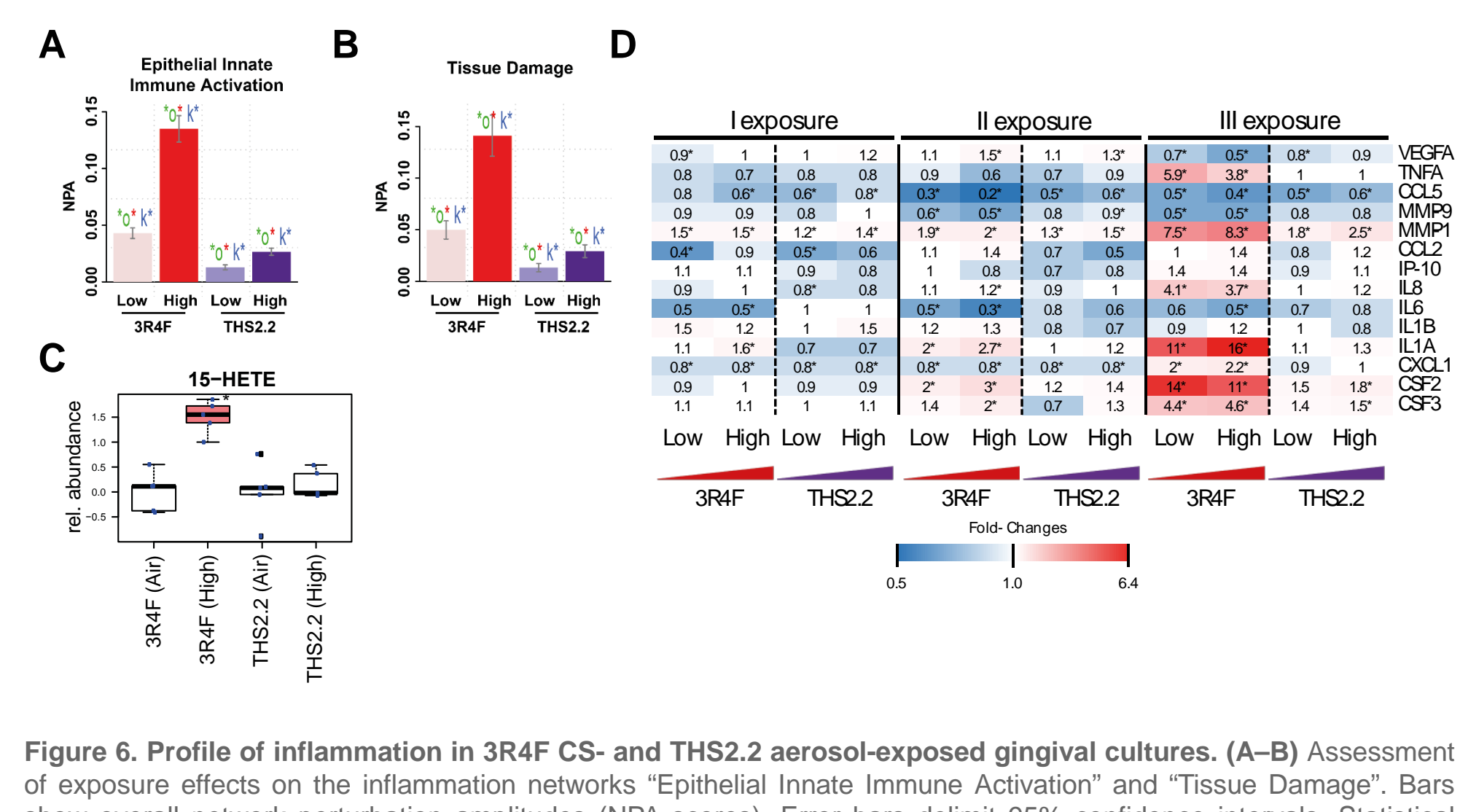


Figure 6. Profile of inflammation in 3R4F CS- and THS2.2 aerosol-exposed gingival cultures. (A-B) Assessment of exposure effects on the inflammation networks “Epithelial Innate Immune Activation” and “Tissue Damage”. Bars show overall network perturbation amplitudes (NPA scores). Error bars delimit 95% confidence intervals. Statistical significance with respect to three different criteria are indicated by colored stars. n=6-9. (C) Metabolomics profiling was conducted 4 h after the 3rd exposure of the tissue to high 3R4F CS and THS2.2 aerosol concentrations. Boxplot summarizes the response of 15-HETE (blue dots indicate individual samples, n=5). Significant differences between exposed groups and their respective sham groups are indicated by filled colored boxes and a star (“***” (FDR <0.05)). (D) Heatmap showing fold-changes of mean concentrations of proinflammatory mediators in exposed cultures relative to their corresponding air controls 24 h after each exposure (I, II, III exposure). Blue and red colors indicate negative or positive fold-changes, respectively, in 3R4F CS- and THS2.2 aerosol-exposed samples compared with air-exposed samples. n=9.

GENE EXPRESSION IMPACT OVERVIEW

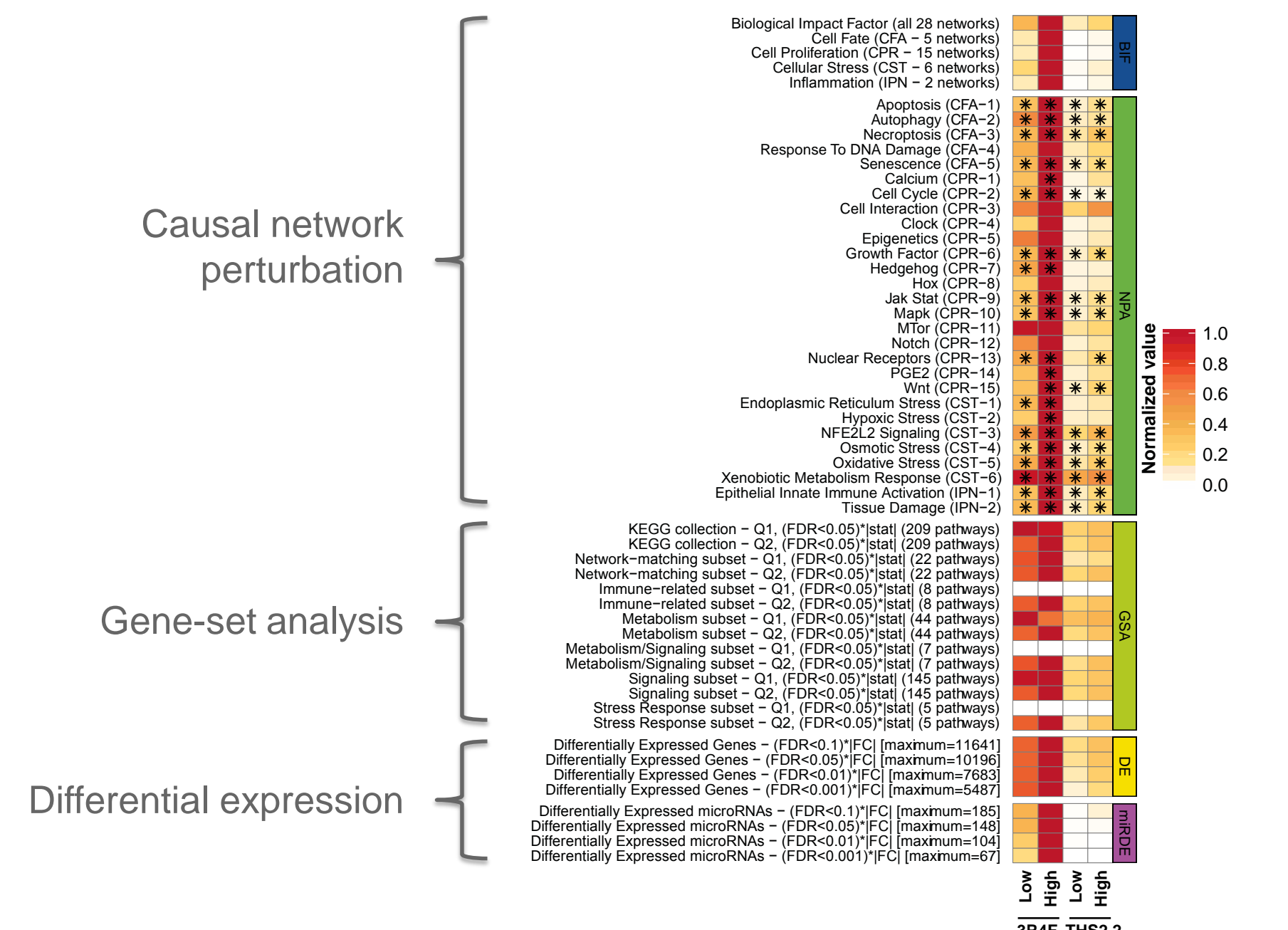


Figure 3. Overview of the impact of 3R4F CS or THS2.2 aerosol exposures on differential expression of genes. Values are normalized to the interval [0, 1] in a row-wise manner. The uppermost panel displays the overall biological impact factor (BIF) and the relative BIFs for four network families (cell fate and angiogenesis (CFA), cell proliferation (CPR), cellular stress (CST), and pulmonary inflammation (IPN)) assessed by a causal network approach [3, 4]. The network perturbation amplitudes (NPA) for the individual networks are shown in the next panel. The “***” indicates statistically significant network perturbations. Overall results of gene set analyses (GSA) are displayed in the next panel for the KEGG collection and the two standard statistical tests (Q1 and Q2). Also shown are specific subsets of the KEGG collection: first, the 22 pathways matching the mechanistic networks, and second, the five broad categories of the 228 pathways contained in the KEGG collection. Displayed values were defined as sums of absolute values of gene set-level statistics (i.e. fold-change mean) for the statistically significant gene sets in each category. The two lower panels show the number of differentially expressed genes (DE) and miRNAs (miRDE) for four distinct statistical significance thresholds, to identify possible threshold effects. Again, sums of absolute values of fold-changes of statistically significant genes or miRNAs are displayed. n=6-9.

XENOBIOTIC METABOLISM

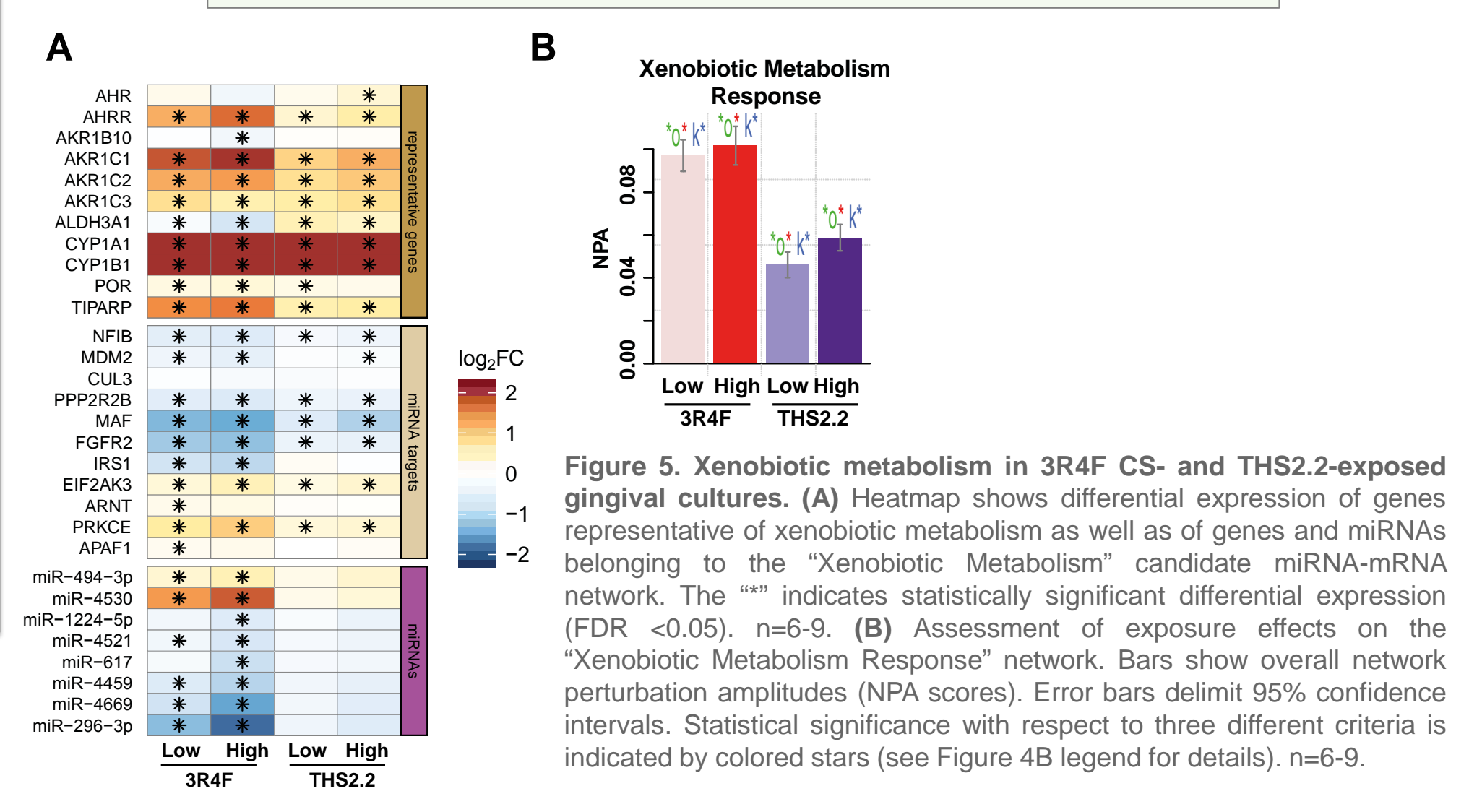


Figure 5. Xenobiotic metabolism in 3R4F CS- and THS2.2-exposed gingival cultures. (A) Heatmap shows differential expression of genes representative of xenobiotic metabolism as well as of genes and miRNAs belonging to the “Xenobiotic Metabolism” candidate miRNA-mRNA network. The “***” indicates statistically significant differential expression (FDR <0.05), n=6-9. (B) Assessment of exposure effects on the “Xenobiotic Metabolism Response” network. Bars show overall network perturbation amplitudes (NPA scores). Error bars delimit 95% confidence intervals. Statistical significance with respect to three different criteria is indicated by colored stars (see Figure 4B legend for details). n=6-9.

KERATINIZATION & CELL ADHESION

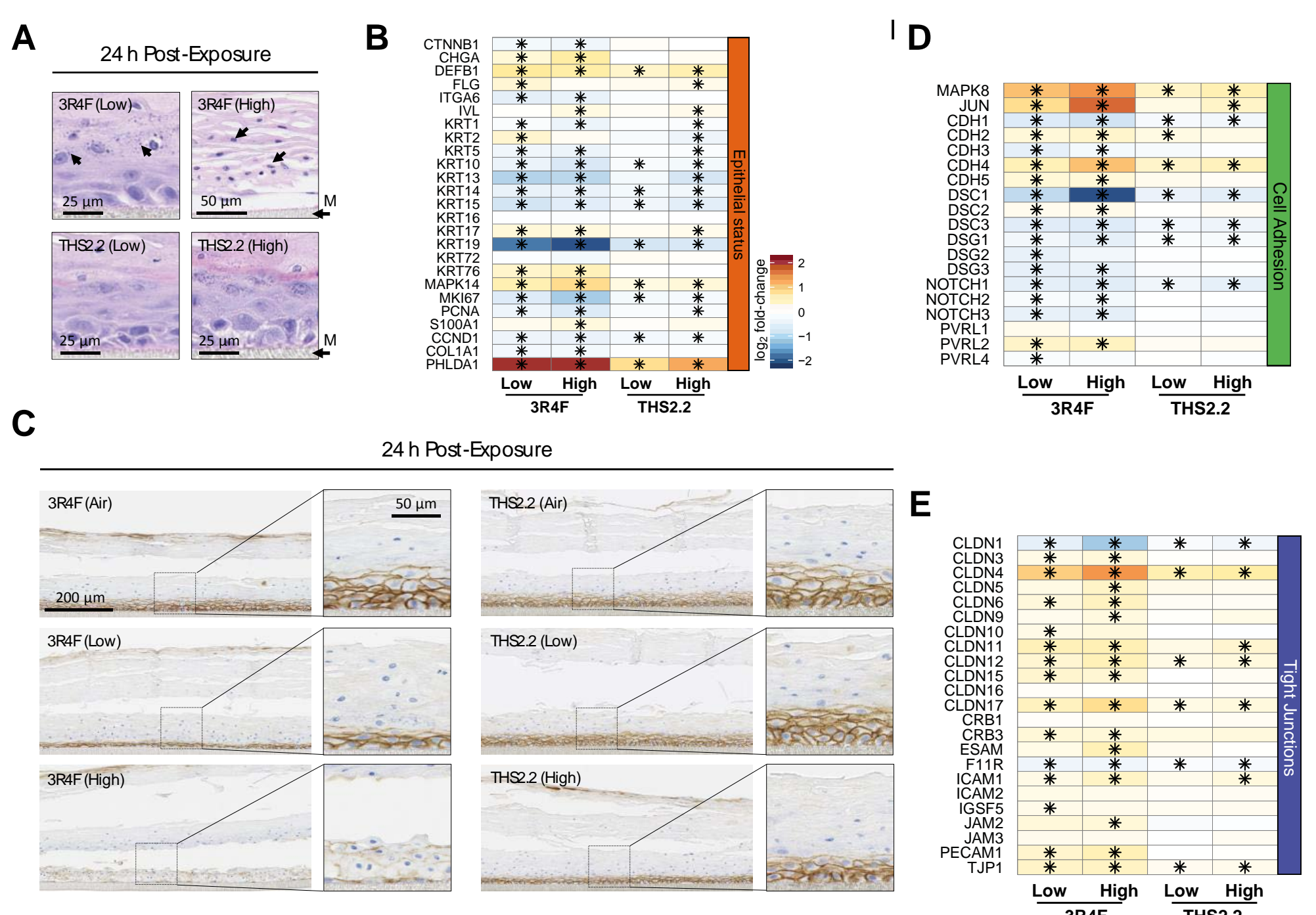


Figure 7. Keratinization and cell-cell adhesion in 3R4F CS- and THS2.2 aerosol-exposed samples. (A) Representative images of H&E-stained gingival culture sections observed 24 h after the last exposure. Arrows indicate keratinocytes (upper left picture) or parakeratosis (upper right picture). Abbreviations: M, membrane. n=9. (B, C) Heatmaps showing differential expression of genes related to epithelial status/cell type, cell adhesion, or tight junctions. Statistical significance (FDR <0.05) is indicated by a star (“***”). n=6-9. (C) Representative images of E-cadherin-stained gingival culture sections observed 24 h after the last exposure. Magnification is set at 20x and 63x for the insets. n=9.

CONCLUSIONS

- Systems toxicology approach was applied for the assessment of THS2.2 aerosol compared to cigarette smoke (CS) on an organotypic gingival epithelium model. Multiple endpoints (e.g., cytotoxicity, transcriptomics, and metabolomics) were combined toward a comprehensive assessment of the exposure effects.
- Major morphological alterations (loss of cell adhesion, keratinization) and cytotoxicity (max. ~30%) were observed after CS exposure but were limited, if not none, upon exposure to THS2.2 aerosol (Figure 2).
- Transcriptomic and metabolomic analysis indicated a general reduction of the impact in THS2.2 aerosol-exposed samples with respect to CS (~79% lower biological impact for the high THS2.2 aerosol concentration compared to CS, and 13 metabolites significantly perturbed upon THS2.2 aerosol exposure vs. 181 for 3R4F CS) (Figure 3 & 4).
- Proinflammatory mediator analysis showed a higher impact in CS-exposed cultures compared to THS2.2 aerosol, with 11 analytes significantly altered by CS vs. 5 by THS2.2 aerosol, showing a reduced fold-change with respect to CS (Figure 6).
- Biological effects induced by CS, such as oxidative stress (Figure 4), xenobiotic metabolism (Figure 5), and inflammation-related processes (Figure 6), are relevant to the pathophysiology of periodontal diseases.
- Overall, THS2.2 aerosol had a statistically significantly lower impact on molecular processes associated with the pathophysiology of human gingival organotypic cultures compared to CS.

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