

# Genomic analysis of L5178Y tk<sup>+/−</sup> cells and their induced tk<sup>−/−</sup> mutant colonies

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## Introduction and Goal

- The genetic basis that underpins the mouse lymphoma assay (MLA) is ostensibly well understood; inactivation of the functional *thymidine kinase* (*tk*<sup>+</sup>) allele in L5178Y cells via mutation or deletion induces trifluorothymidine (TFT) resistance and *tk*<sup>−/−</sup> mutants can be selected for against a background of *tk*<sup>+/−</sup> wild-type cells via TFT-mediated enrichment (Clements, 2000).
- However, despite its widespread use over the past 20 years for hazard identification purposes, few studies have sought to characterise in greater detail the colonies of cells which propagate as a result of mutagenesis in the assay.
- The advent of whole genome sequencing has the potential to enhance mutagenicity testing since genetic changes can now be investigated with high level of detail at the genome level.
- Next generation sequencing (NGS) technology was used to sequence the genomes of:
  - an L5178Y cell line frequently used in MLA studies.
  - tk*<sup>−/−</sup> mutant colonies induced by two prototypical mutagens in the assay.
- The goal of this pilot study was to exploit NGS technology with a view to shedding new light on the genome of cells used in the MLA, as well as the changes induced as a result of the exposure to mutagens.

## Materials and Methods

- Cell line:** L5178Y *tk*<sup>+/−</sup> clone 3.7.2C IVGT (Public Health England, UK). Spontaneously-occurring *tk*<sup>−/−</sup> mutant cells were purged from the population via a cleansing procedure that used methotrexate to select against *tk*-deficient cells and thymidine, hypoxanthine and guanine to ensure optimal growth of *tk*-proficient cells.
- MLA:** carried out according to established standards (Moore *et al.*, 2006). Briefly, cells (10<sup>6</sup>) were exposed to either methyl methanesulfonate (MMS; 10 and 20 µg/ml) in the absence of S9 or 7,12-dimethylbenz[*a*]anthracene (DMBA; 1 and 1.5 µg/ml) in the presence of S9 for 4 hours in a shaking incubator set at 37°C. DMSO was the solvent control in each case.
- MLA procedure:**
- tk*<sup>−/−</sup> mutant colony harvesting:** following MF and ISC determination, small and large *tk*<sup>−/−</sup> mutant colonies were randomly selected and removed from the wells for processing.
- DNA extraction and sequencing:** Colonies were processed via the QiaAMP® DNA mini kit (Qiagen, USA). Genomic DNA extracted from colonies was processed via the Ovation® Ultralow DR Multiplex System kit (Nugen, USA). All libraries were sequenced on an Illumina HiSeq-2500 with runs of 2 × 100 bases. Average sequencing depth for the L5178Y cell line was 58-fold and between 8 to 31-fold for the mutants.
- Bioinformatics:** The paired-ends were trimmed and cleaned using Trimmomatic (Bolger *et al.*, 2014) and aligned to the GRCh38 version of the mouse genome using the BWA MEM (Li H, 2013) algorithm. SNPs were called using the software FreeBayes (Garrison E & Marth G, 2012):
  - L5178Y heterozygous sites:** Stringent quality filtering was applied to identify reference heterozygous sites in the L5178Y cell line.
  - Loss of heterozygosity:** Heterozygosity at the reference sites in the mutants was determined by running FreeBayes with very permissive parameters, so as to avoid false negative heterozygosity calls.

## Heterozygous point mutations in L5178Y

Table ①

Subset	All chromosomes	Chromosome 11
Total	2,078,744	221,108
Shared with Mouse Genome Project	1,803,902	205,952
Exclusive to L5178Y	275,842	15,156

Table ②

Gene	Reference amino acid	Position	Mutant amino acid	SIFT score	Gene function
Asf1a	D	57	N	0.0006	Histone chaperone ASF1A
Ino80	S	270	P	0.0176	Chromatin remodeling complex
Fbxo6	R	76	K	0.028	Putative control of the cell cycle
Brca2	N	737	T	0.0424	DNA repair associated
Rfc3	R	372	H	0.0023	Replication factor C
Lig1	Y	671	H	0.0252	DNA ligase
Herc2	R	1744	L	0.0028	Retention of repair proteins on damaged chromosomes

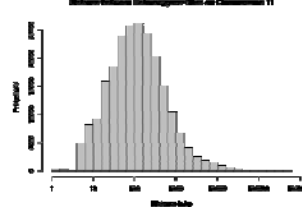
### Survey of heterozygous sites in L5178Y:

Table ①: Survey of SNPs located in the L5178Y genome.

Table ②: Mutations exclusive to L5178Y, which are predicted to impact DNA repair enzymes, according to the SIFT algorithm (Ng P & Henikoff S, 2001).

## Heterozygosity on chromosome 11

Figure ①

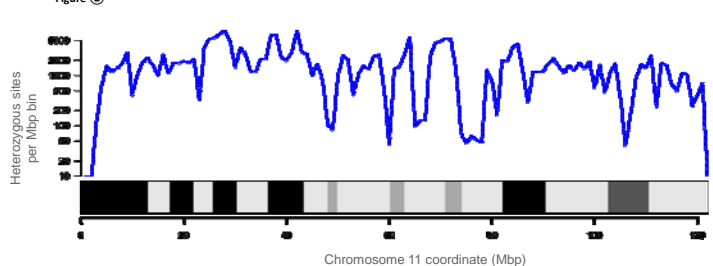


### Survey of heterozygous sites in L5178Y:

Figure ①: The distance between heterozygous sites on chromosome 11 limits the resolution with which chromosome loss can be determined

Figure ②: Number of heterozygous sites per 1 Mbp bin on chromosome 11

Figure ②



## Mapping loss of heterozygosity on chromosome 11

Figure ③

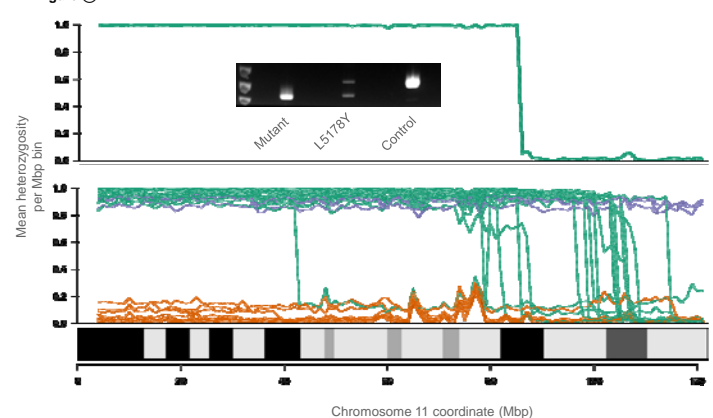


Figure ③: Using heterozygous sites in L5178Y to map loss of heterozygosity in mutant colonies

- Upper Panel:** Example of abrupt loss of heterozygosity (LOH) due to breakage. Inset: Confirmation of LOH by PCR with Agl2 primers (Liechty *et al.* 1996)
- Lower Panel:** LOH mapping for all colonies; mauve - no LOH; green - LOH; orange - loss of whole chromosome.

## Summary and Conclusion

- Analysis of the L5178Y *tk*<sup>+/−</sup> cell line's genome revealed numerous pre-existing small polymorphisms, many of which are shared with various mouse strains.
- Several novel mutations in L5178Y were identified, which could explain the sensitivity of this cell line to mutation.
- Loss of heterozygosity at pre-existing heterozygous sites can be observed in many mutant colonies by NGS sequencing.
- Mapping LOH to allow breakpoints on chromosome 11 to be identified, and the density of heterozygous sites along the chromosome defines the resolution of this method.
- In conclusion, NGS-mediated genomic analysis of *tk*<sup>−/−</sup> mutant cells may shed new light on the genetic basis of the MLA, while also potentially providing additional evidence for the risk assessment of mutagenic compounds.

## References

- Clements J. 2000, The mouse lymphoma assay, *Mutat. Res.* 455: 97-110.
- Moore M *et al.*, 2006, Mouse Lymphoma Thymidine Kinase GeneMutation Assay: Follow-up Meeting of the International Workshop on Genotoxicity Testing-Aberdeen, Scotland, 2003- Assay Acceptance Criteria, Positive Controls, and Data Evaluation, *Environ. Mol. Mutagen.* 47: 1-5.

